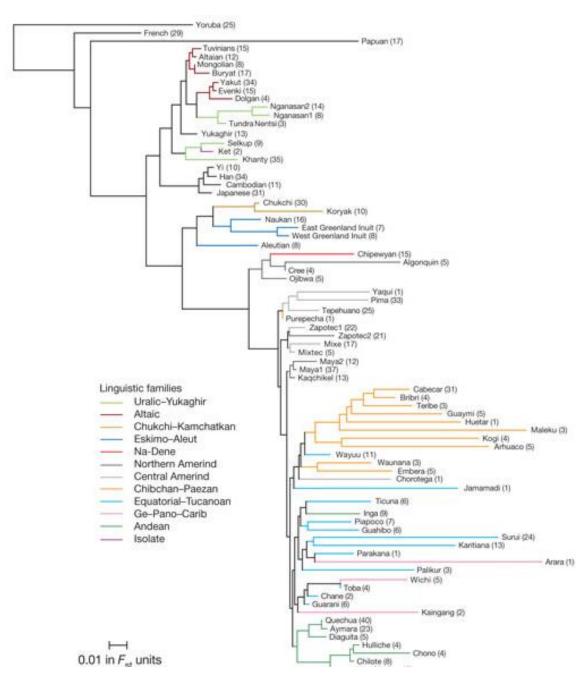
# Multiple sequence alignments similarity without sequence similarity

Andrew Torda, Bioinformatics, Sommersemester 2013

Bis Jetzt

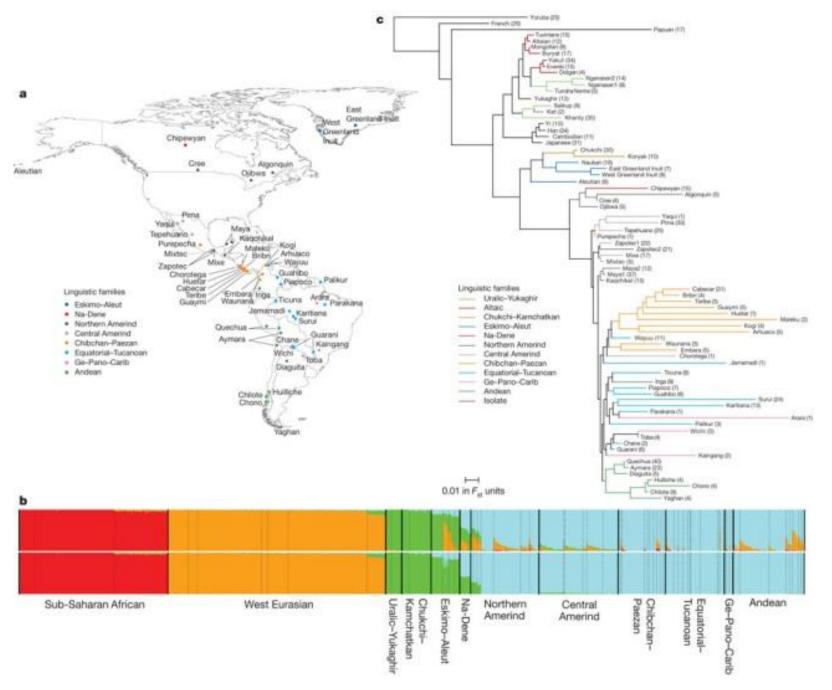
- Man hat eine Sequenz (Protein oder Nukleotid)
- Man will so viel wie möglich finden, um
  - Struktur vorherzusagen
  - Funktion vorherzusagen
- Jetzt Alignments, Evolution & Funktion



### 52 Native American groups

Reich, D., ...Ruiz-Linares, A., Nature, 488, 370 (2012), Reconstructing Native American .. History

07/05/2013 [2]



Reich, D., ...Ruiz-Linares, A., Nature, 488, 370 (2012), Reconstructing Native American .. History

## **Multiple alignments**

...

...

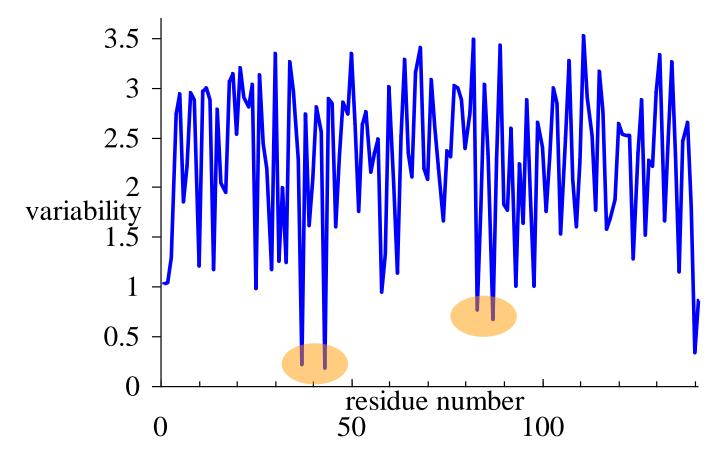
... ...

- what does a set of sequences look like?
- data for a ullethaemoglobin
- summarise this data

VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG  $mostly \ for \ proteins^{\text{vlspadktnvkaawgkvgahageygaealermflsfpttktyfphfdlshgsaqvkghg}}_{\text{wlspadktnvkaawgkvgahageygaealermflsfpttktyfphfdlshgsaqvkghg}}$ VLSPADKTNVKAAWGKVGAHAGEYGAEALEKMFLSFPTTKTYFPHFDLSHGSAQVKGHG LSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKTNVKAAWGKVGAHAGDYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPDDKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKTHVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKTNVKAAWGKVGAHAGEYGAEAWERMFLSFPTTKTYFPHFDLSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEAWERMFLSFPTTKTYFPHFDLSHGSAOVKGHG VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAOVKGHG VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSAADKTNVKAAWSKVGGHAGEYGAEALERMFLGFPTTKTYFPHFDLSHGSAQVKAHG VLSAADKTNVKAAWSKVGGHAGEYGAEALERMFLGFPTTKTYFPHFDLSHGSAOVKAHG VLSADDKANIKAAWGKIGGHGAEYGAEALERMFCSFPTTKTYFPHFDVSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEAFERMFLSFPTTKTYFPHFDLSHGSAQVKGQG VLSPADKTNVKAAWGKVGAHAGEYGAEAFERMFLSFPTTKTYFPHFDLSHGSAQVKGQA VLSAADKSNVKAAWGKVGGNAGAYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAOVKGHG VLSPADKSNVKATWDKIGSHAGEYGGEALERTFASFPTTKTYFPHFDLSPGSAQVKAHG VLSPADKSNVKAAWGKVGGHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTGTYFPHFDLSHGSAQVKGHG VLSSADKNNVKACWGKIGSHAGEYGAEALERTFCSFPTTKTYFPHFDLSHGSAQVQAHG VLSAADKSNVKAAWGKVGGNAGAYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSANDKSNVKAAWGKVGNHAPEYGAEALERMFLSFPTTKTYFPHFDLSHGSSOVKAHG VLSPADKSNVKAAWGKVGGHAGDYGAEALERMFLSFPTTKTYFPHFDLSHGSAOVKGHG

### **Conservation / variability**

Look at residues 37, 43, 83 and 87



- how do we get these and what does it mean?
- what does it mean for this protein ?

07/05/2013 [5]

### **Conserved residues**

Proximity to haem group

• green residues

• more on pro 37 later

### **Beliefs in multiple sequence alignments**

Similar proteins found in many organisms

- rarely identical
- where they are conserved will be connected with function
- how much they vary will reflect evolution (phylogeny)
  How many homologues might you have ?
- many
  - some DNA replication proteins almost every form of life
  - some glycolysis proteins from bacteria to man
  - .
- few
  - some exotic viral proteins
  - some messengers exclusively in human biochemistry

### Many sequences - rigorous alignment

- two sequence alignment
  - optimal path through  $n \times m$  matrix
- three sequence alignment
  - optimal path through  $n \times m \times p$  matrix
- four sequence alignment
  - ...
- *m* sequence alignment of *n* residues....  $O(n^m)$

Excuse to use lots of approximations

• no guarantee of perfect answer

Reasonable starting point

• begin with pairs of proteins

### **Scoring schemes**

$$S_{a,b} = \sum_{i=1}^{N_{res}} \operatorname{match}(s_{a,i}, s_{b,i})$$

In pairwise problem

VLSPADKSNVKAGWGQVGAHAGDYGAEAIERMYLSFPSTKTYFPHTDISHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG

- Sum over match() where *N<sub>res</sub>* is sequence length
- match(s<sub>a,i</sub>, s<sub>b,i</sub>) is the match/mismatch score of sequence a and b at position i
- invent a distance between two sequences like

$$d_{a,b} = 1 - \frac{S_{a,b}}{100 \cdot N_{res}}$$
 or  $d_{a,b} = \frac{1}{S_{a,b}}$ 

 distance measure –which sequences are most similar to each other

## Scoring schemes for a multiple alignment

In the best alignment

- 1 is aligned to 2, 3, ..
- 2 to 3,4, ...

VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG
 VITP-EQSNVKAAWGKVGAHAGEYGAEALEQMFLSYPTTKTYFP-FDLSHGSAQIKGHG
 MLSPGDKTQVQAGFGRVGAHAG--GAEALDRMFLSFPTTKSFFPYFELTHGSAQVKGHG
 VLSPAEKTNIKAAWGKVGAHAGEYGAEALEKMF-SYPSTKTYFPHFDISHATAQ-KGHG
 VTPGDKTNLQAGW-KIGAHAGEYGAEALDRMFLSFPTTK-YFPHYNLSHGSAQVKGHG
 VLSPAEKTNVKAAWGRVGAHAGDYGAEALERMFLSFPSTQTYFPHFDLS-GSAQVQAHA
 VLSPDDKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG

### Mission: for $N_{seq}$ sequences

•  $S_{ab}$  : alignment score sequences a and b

$$score = \sum_{b \neq a}^{N_{seq}} \sum_{a=1}^{N_{seq}} S_{a,b}$$

- not quite possible
  - if I move sequences 4 and 5, may make a mess of 5 and 2

## Aligning average sequences

VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VITPAEKTNVKAAWGKVGAHAGEYGAEALEQMFLSYPTTKTYFPHFDLSHGSAQIKGHG

### and

IITPGDKTNVKAAFGKVGAHGGEYGAEALDRMFISFPSTKTYYPHFDLSHASAQVKAHG VITPAEQTNIKGAWGQIGAHAGDYAADALEQMFLSYPTSKTYFPYFDLTHGSAQIKGHG VITPAEKTQVKAAWGKVGGHAGEYGAEAIEQMFLTYPTTQTYFPHFELSHGTAQIKGHG

- At each position
- use some kind of average in scoring
- if a column has  $2 \times D$  and  $1 \times E$  score
  - score as 2/3 D + 1/3 E
- later.. call the average of S1 and S2: av(S1, S2)

## Summarise ingredients

- pairwise scores + distances
- ability to align little groups of sequences

### **Progressive alignments**

Known as guide tree / progressive / neighbour joining method

Steps

- build a distance matrix
- build a guide tree
- build up overall alignment in pieces

### **Progressive alignment - tree**

- S1 ATCTCGAGA
- S2 ATCCGAGA
- S3 ATGTCGACGA
- S4 ATGTCGACAGA
- S5 ATTCAACGA

S1 calculate the S2 .11 distance matrix S3 .20 .30 .27 S4.36 .09 .30 .33 S5 .23 .27 S2 S1 S3 S4 S5 S1 calculate guide tree S2 S3 S4 S5

Compute pairwise alignments,

### Multiple alignment from guide tree

- gaps at early stages remain
- problems..
- S1/S2 and S3/S4 good
  - no guarantee of S1/S4 or S2/S3

• av(S1,S2) is average of S1 and S2

#### align S1 with S2

S1	ATCTCGAGA
S2	ATC-CGAGA

### align S3 with S4 S3 ATGTCGAC-GA S4 ATGTCGACAGA

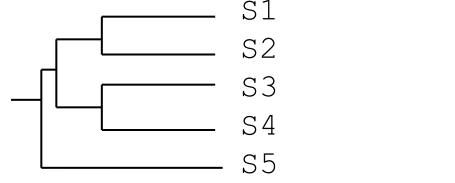
### align av(S1,S2) with av(S3,S4)

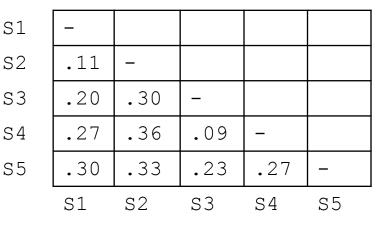
S1	ATCTCGAGA
S2	ATC-CGAGA
S3	ATGTCGAC-GA
S4	ATGTCGACAGA

### align av(S1,S2,S3,S4) with S5

- S1 ATCTCGA--GA
- S2 ATC-CGA--GA
- S3 ATGTCGAC-GA
- S4 ATGTCGACAGA
- S5 AT-TCAAC-GA

### **Problems and variations**

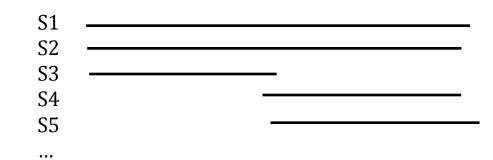




What order should we join ?

- pairs are easy (S1+S2) and (S3+S4)
- which next?

Real breakdown



- S1 and S2 are multi-domain proteins
  - S3 is not really related to S4 or S5
  - distance matrix elements are rubbish

### Given an alignment

How reliable / believable ?

- set of very related proteins (an enzyme from 100 mammals)
  - no problem
- diverse proteins (an enzyme 100 organisms, bacteria to man)
  - maybe lots of little errors
- can break completely (domain example)

Is the tree a "phylogeny" ? A reflection of evolution ?

• more later

### **Measuring conservation / entropy**

Gibbs entropy

- how much disorder do I have ?  $S = -k \sum_{i=1}^{N_{states}} p_i \ln p_i$
- in how many states may I find the system ? Our question
- look at a column how much disorder is there ?

VLSPADKTNVKAAWGKVGAFA GEYGAEALERMFLSFPTTKTYFPHED LSHGSAQVKGHG VITP-EQSNVKAAWGKVGAFA GEYGAEAIEQMFLSYPTTKTYFPHED LSHGSAQIKGHG MLSPGDKTQVQAGFGRVGAFA G--GAEAVDRMFLSFPTTKSFFPYFE LTHGSAQVKGHG VLSPAEKTNIKAAWGKVGAFA GEYGAEAAEKMF-SYPSTKTYFPHED ISHATAQ-KGHG -VTPGDKTNLQAGW-KIGAFA GEYGAEALDRMFLSFPTTK-YFPHY N LSHGSAQVKGHG VLSPAEKTNVKAAWGRVGAFA GEYGAEALERMFLSFPSTQTYFPHED LS-GSAQVKGHG

> no disorder

much disorder

Calculate an "entropy" for each column

## Entropy

- forget k (Boltzmann just scaling)  $S = -\sum_{i=1}^{N_{states}} p_i \ln p_i$ We have a protein
- 20 possible states

What if a residue is always conserved ?

 $S = \ln(1) = 0$  (no entropy)

What if all residues are equally likely ?  $p_i = \frac{1}{20}$ 

$$S = -\sum_{i=1}^{20} \frac{1}{20} \ln \frac{1}{20} = -20 \cdot \frac{1}{20} \ln \frac{1}{20}$$

**≈** 3

• my toy alignment...

07/05/2013 [18]

## Entropy

## First column is boring

### Second

$$p_{\rm D} = {}^{5}\!/_{7}$$
  
 $p_{\rm E} = {}^{1}\!/_{7}$ 

 $p_{\rm N} = 1/_{7}$ 

$$S = -\left(\frac{5}{7}\ln\frac{5}{7} + \frac{1}{7}\ln\frac{1}{7} + \frac{1}{7}\ln\frac{1}{7}\right)$$
  
\$\approx 0.8\$

• example from start of this topic

### **Entropy from DNA**

Exactly as for proteins Will numbers be larger or smaller ?

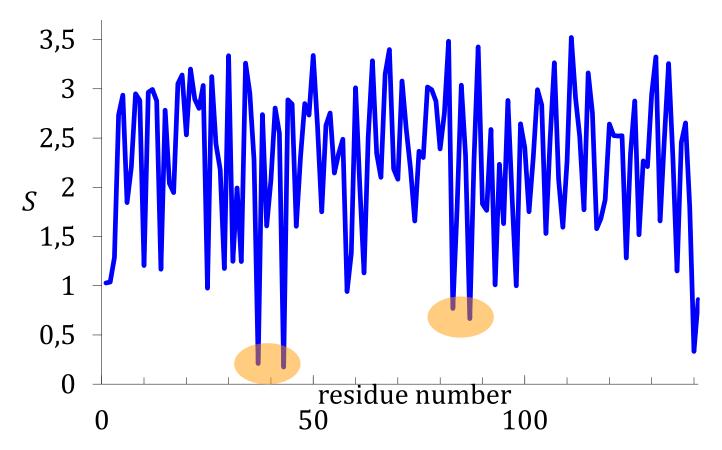
max possible entropy

$$S = -4\left(\frac{1}{4}\ln\frac{1}{4}\right)$$
$$= -\ln\frac{1}{4}$$

 $\approx 1.4$ 

### Haemoglobin conservation

Look at residues 37, 43, 83 and 87



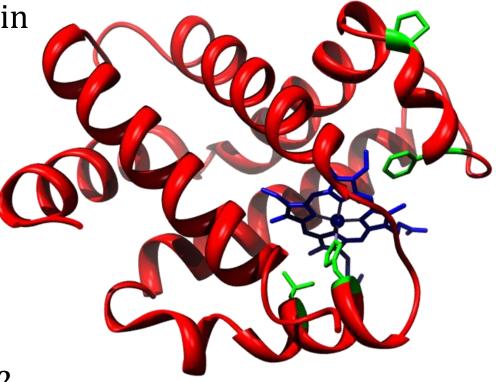
4 residues (maybe more) stand out as conserved

• why ?

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### **Conserved residues in haemoglobin**

- 3 of the sites are easy to explain
- interact with haem group
- Look at fourth site
- proline
- end of a helix



What is special about proline ?

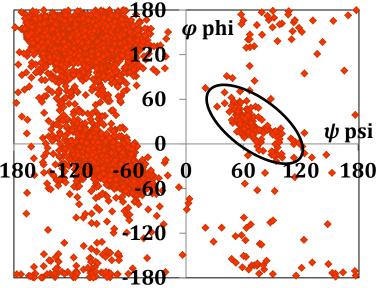
- no Hbond donor
- here if it mutates, maybe haemoglobin does not fold

### **Conservation for structure**

Some residues have very special structural roles

- proline not an H-bond donor
  - often end of a helix
- glycine can visit part of  $\varphi \psi$  plot
  - found in some turns
- Are all gly residues so important?
- NO they occur in many places sometimes in turns

Are all pro residues very conserved ? No



### **Conservation for function**

In a serine protease

- always a "catalytic serine"
- can it mutate ? Not often

In haemoglobin – residues necessary for binding haem

- can they mutate ? rarely
- changes properties of haemoglobin (bad news)

Dogma

residues in active site will be more conserved than other sites

### **Important summary**

Conservation may reflect

- important function
- structural role
- Mutagenesis / chemistry
- what residue may I change to allow binding to a solid substrate ? (for biosensor/immobilized enzyme ?)
- I want to try error prone PCR to select for new enzyme activity which sites might I start with (active site) ?
- Drug design example
- target is an essential protein (basic metabolism, DNA synthesis, protein synthesis..)
- is there some set of sequence features common to pathogen, different to mammalian protein ?

### **Evolution – do not trust conservation**

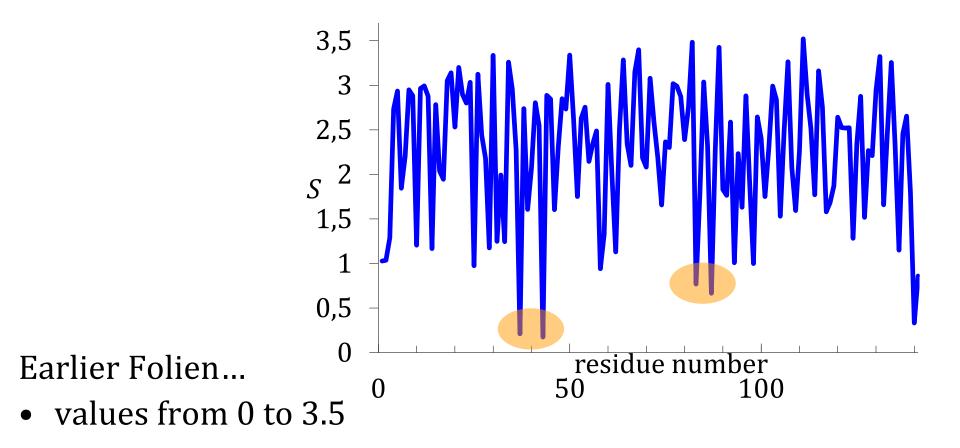
Imagine: two possible systems for some important enzyme

- 1. active site fits to essential biochemistry
  - any mutation you lose
  - you see active site residues as conserved in a conservation plot
- 2. maybe enzyme is not absolutely perfect
  - some mutations kill you
  - some mutations OK
  - site does not appear perfectly conserved

Where would you evolve to ?

- 1. very fragile
- 2. likely to survive mutations

### **Conservation – how meaningful ?**



What if I used more homologues ?

### **Conservation – how meaningful ?**

Example sequence (1ab4, DNA gyrase)

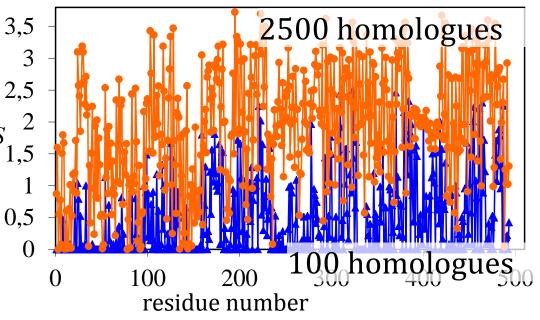
find 100 close homologues (mostly > 80% similarity)
 – calculate conservation

S

 find 2 500 close homologues (mostly > 50 % similarity) calculate conservation

Fewer sequences

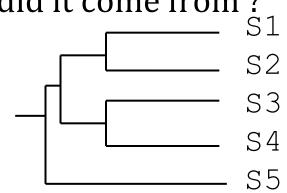
- lots of conserved sites
- you can get the answer 0, you want



## **Phylogeny / Evolution**

Purely academic ? For fun ? Not always

- possibly useful in explaining disease propagation
  - where did HIV come from ?
  - where did the flu pandemics come from ?
  - virus infects banana crop where did it come from ?
- previously we had a "guide tree"
  - did (S1,S2) and (S3,S4) share an ancestor but not S5 ?
  - not so good
- branch lengths do not reflect evolutionary time
- there may be other similar trees which could be evolutionary paths



### **Evolutionary time**

Compare two DNA sequences see

- 1 mutation (represents time *t*)
- 2 mutations (time 2*t*)
- 3 mutations (time 3*t*)...
- No !

### After some evolution

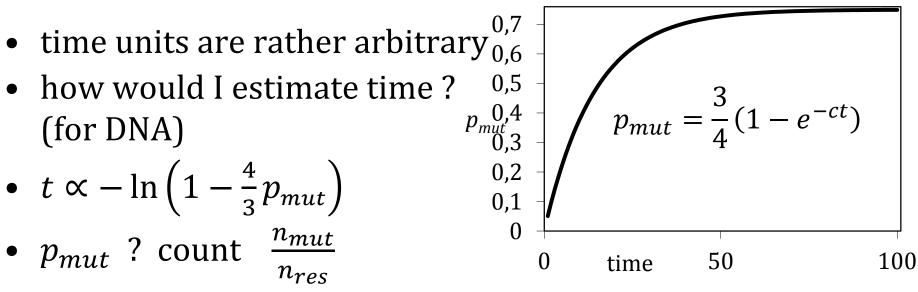
А	$\rightarrow C$	$\rightarrow$	G	two ever	nts (although looks like $A \rightarrow G$ )
А	$\rightarrow C$	$\rightarrow$	G	$\rightarrow$ C $\rightarrow$ A	looks like zero mutations

If I have infinite time

- all bases / residues equally likely
- $p_{mut} = 3/4 = 0.75$  (DNA) or  $p_{mut} = 19/20$  (protein)

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### **Mutation probability**



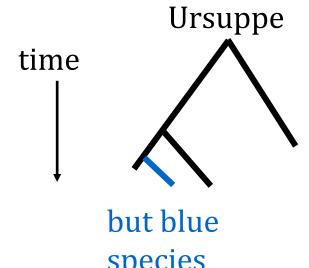
• scaling of *t* not so important (relative time)

For short times,  $p_{mut}$  changes fast

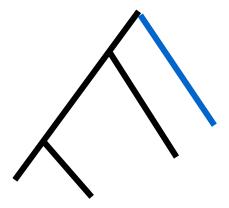
- for small *t*, distances will be more reliable
  - as will be alignments
- Is this enough for phylogeny ?
- what about reliability ?

## **Problems in phylogeny**

- not all sites mutate equally quickly
- not all species mutate equally quickly



but blue species (protein) mutates quickly



blue appears to have branched off earlier

### **Problems estimating time**

- 1. mutation rates vary wildly
  - changing environments pH, temperature,..
- 2. imagine time *t* is such that  $p_{mut}$  = 0.25
  - we have random events
  - sometimes you see 23% mutation, sometimes 28%
- time estimates will never be accurate
- maybe we cannot find the correct tree
  - can we roughly estimate reliability?

## Reliability

# Think of first alignment

VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VITP-EQSNVKAAWGKVGAHAGEYGAEAIEQMFLSYPTTKTYFP-FDLSHGSAQIKGHG MLSPGDKTQVQAGFGRVGAHAG--GAEAVDRMFLSFPTTKSFFPYFELTHGSAQVKGHG VLSPAEKTNIKAAWGKVGAHAGEYGAEAAEKMF-SYPSTKTYFPHFDISHATAQ-KGHG -VTPGDKTNLQAGW-KIGAHAGEYGAEALDRMFLSFPTTK-YFPHYNLSHGSAQVKGHG VLSPAEKTNVKAAWGRVGAHAGDYGAEAGERMFLSFPSTQTYFPHFDLS-GSAQVQAHA VLSPDDKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG

What would happen if you deleted a column ?

- if the data is robust /reliable
  - not much
- if the tree is very fragile /sensitive
  - tree will change

better...

### Reliability

Repeat  $10^2$  to  $10^3$  times

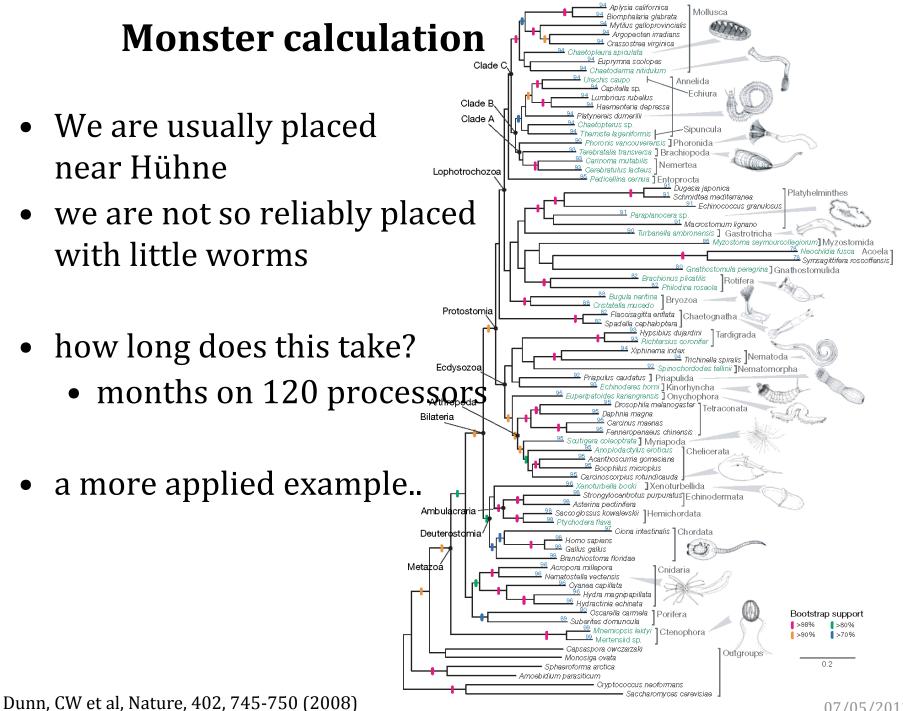
- delete 5 to 10 % of columns
- copy random columns so as to have original size
- recalculate tree

How often did you see each branch?

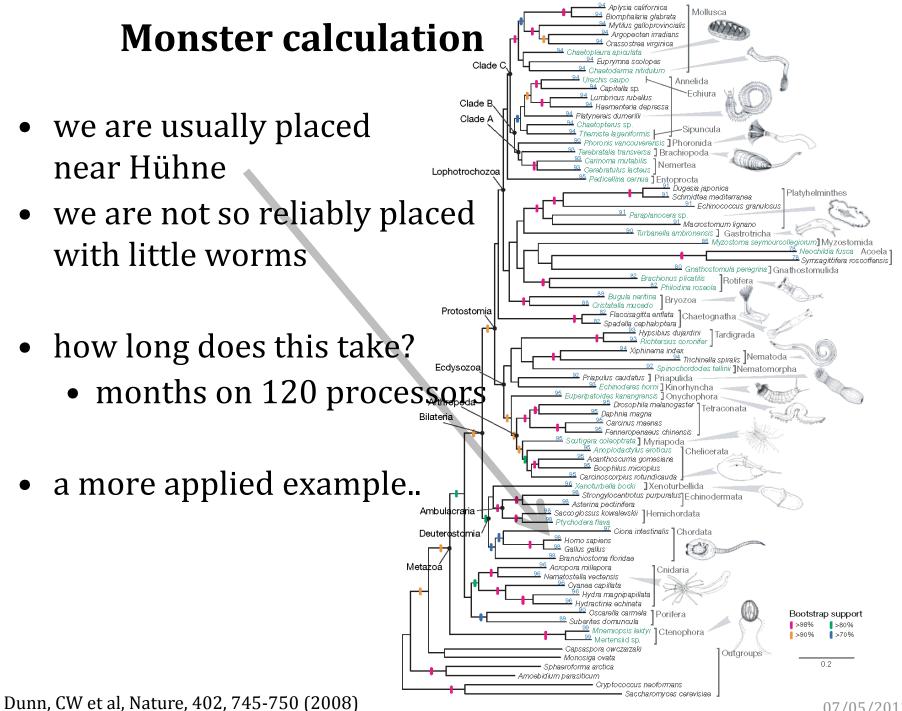
VLSPADKTNVKAAWGKVGAHAGE /GAEALERMFLSFPT FKTYFPHFDLSHGSAQVF GHG VITP-EQSNVKAAWGKVGAHAGE /GAEAIEQMFLSYPT FKTYFP-FDLSHGSAQIF GHG MLSPGDKTQVQAGFGRVGAHAG--GAEAVDRMFLSFPT FKSFFPYFELTHGSAQVF GHG VLSPAEKTNIKAAWGKVGAHAGE /GAEAAEKMF-SYPS FKTYFPHFDISHATAQ-FGHG -VTPGDKTNLQAGW-KIGAHAGE /GAEALDRMFLSFPT FK-YFPHYNLSHGSAQVF GHG VLSPAEKTNVKAAWGRVGAHAGE /GAEALERMFLSFPS FQTYFPHFDLS-GSAQVCAHA

### Monster example

- generate lots of trees
- for each subtree
  - see how often it is present
- example from cover of nature

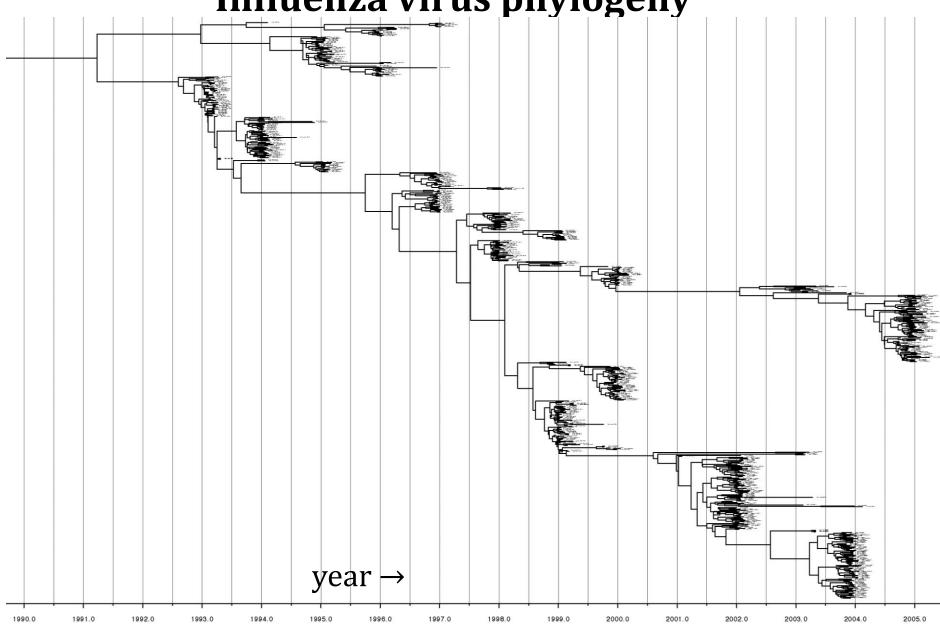


07/05/2013 [38]



07/05/2013 [39]

#### Influenza virus phylogeny



Rambaut, A., .. Holmes, C. The genomic.. influenza A virus, Nature 452, 1-6, 2008

07/05/2013 [40]

# **DNA or protein sequences ?**

- regulatory regions, RNA genes
- synonymous mutations
  - more common than ones that change amino acids
- non-synonymous mutations (amino acid changes)
  - more information  $D \rightleftharpoons E$ ,  $I \rightleftharpoons L \rightleftharpoons V$ , ...
- frame shifts
  - destroy an amino acid sequence
  - small change at DNA level
    - how often do they occur ?
- alignment reliability
  - proteins
    - better, amino acid similarity
  - DNA
    - less information

# **DNA or protein sequences ?**

	protein	DNA	time
synonymous changes	no	yes	short
a.a. changes	yes	no	longer
frame shifts	no	yes	
non-coding regions	no	yes	

Short time

• use DNA

#### Longer time

• use proteins

# Summary

- multiple sequence alignment conservation
  - find important residues (function or structure)
  - can quantify conservation
- relations between most similar proteins are most reliable
- best tree is never found
  - too difficult algorithmically
  - lots of errors evolution is a random process
- rough idea of reliability
- quick tree possible for hundreds of sequences
- more complicated methods only practical for smaller numbers of sequences

### **Protein structures and comparisons**

Ultimate aim

- how to find out the most about a protein
- what you can get from sequence and structure information

On the way..

- remote similarities between proteins
- sequence versus structural similarity
- Detour
  - protein coordinates representation, accuracy
- measures for similarity of coordinates

# **Sequence and structure similarity**

Claim from before

 if two sequences are similar – they are related – structures are similar

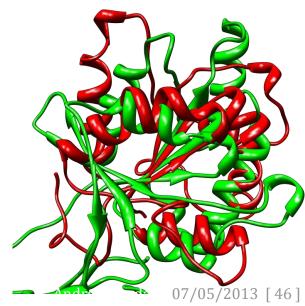
Question

• if two sequences are different are their structures different?

### **Remote similarities**

1cbl & 1eca (haemoglobin & erythrocruorin) 14 % sequence id

1fyv & 1udx, TLR receptor and nucleotide binder, 9 % sequence id



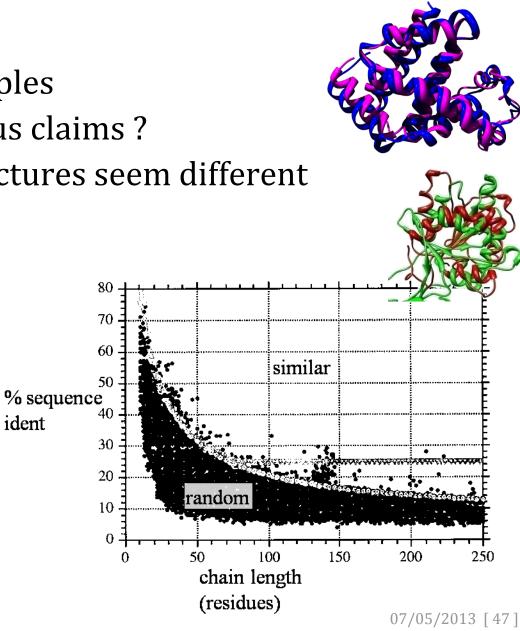
# No sequence similarity – similar structures

Are these rare ?

- easy to find 100s of examples
  Does this agree with previous claims ?
- dot in diagram two structures seem different

If sequences are similar

- structures will be similar
- If sequences are different
- one does not know



Rost, B.Prot Eng, 12,85-94, 1999

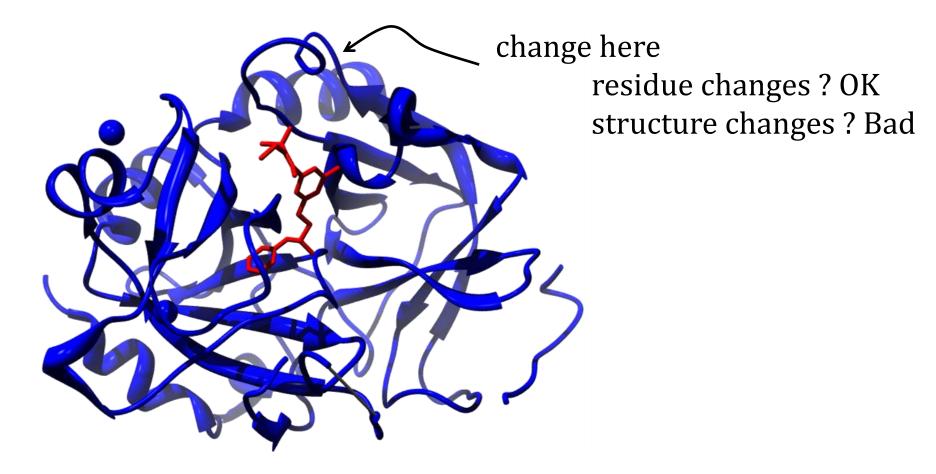
### **Structure versus sequence similarity**

Clear statement

- sequence changes faster than structure
  Reason ? Unclear
- possibility..
- protein function depends on having groups in orientation in space

# Why can sequence change

View of molecular evolution...



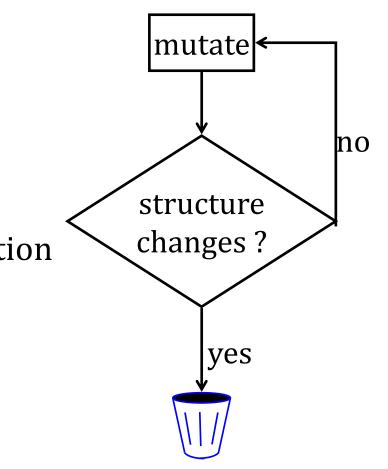
# Simple view of molecular evolution

mutate continuously

- mutations which are not lethal
  - may be passed on (fixed)
- if structure changes
  - protein probably will not function
  - not passed on

Result

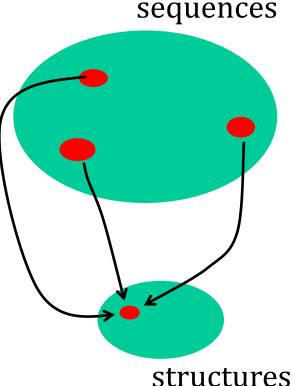
- evolution will find many sequences
  - compatible with structure
  - compatible with function
- how else would we see this ?



### **Sequence vs structure evolution**

Sayings..

- Sequence and structure space
  - sequence space is larger
    - many different sequences map to similar structure
- sequence evolves faster than structure



• Truths...

# **Practical Consequences**

Sequences of proteins are nearly always known

- similar sequence
  - usually similar structure, similar function
- sequences not (obviously) related
  - maybe similar structure
  - maybe similar function

What if structures are known?

# **Sequence vs structure similarity**

When comparing proteins

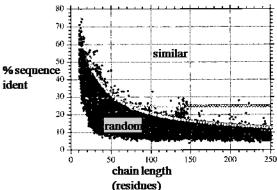
more information is always better (sequence, structure,function)

Similar sequences

- structure and function will be similar
  - remember threshold graphs from earlier

Similar structures, different sequences

- evolutionary relationship implied but
  - bigger evolutionary distance
- not enough to be confident about function
- what do we mean by similar structures ?



### **Comparing protein structures**

Comparing protein structures

- not an O(n),  $O(n^2)$ ,  $O(n^3)$  problem
- requires approximations
  - optimal answer never guaranteed

# **Summarise and Stop**

Multiple sequence alignments

- for conservation
- for phylogenies

Phylogenies

• not as reliable as the pictures imply

Structure vs sequence evolution

- sequence changes faster
- sequence similarity means a closer evolutionary relationship
  - functional similarity