# Multiple sequence alignments similarity without sequence similarity

Andrew Torda, Bioinformatics, Sommersemester 2017

Bis jetzt

- Man hat eine Sequenz (Protein oder Nukleotid)
- Man will so viel wie möglich finden, um
  - Struktur vorherzusagen
  - Funktion vorherzusagen
- Jetzt Alignments, Evolution & Funktion

# **Multiple alignments**

...

... ...

- what does a set of sequences look like?

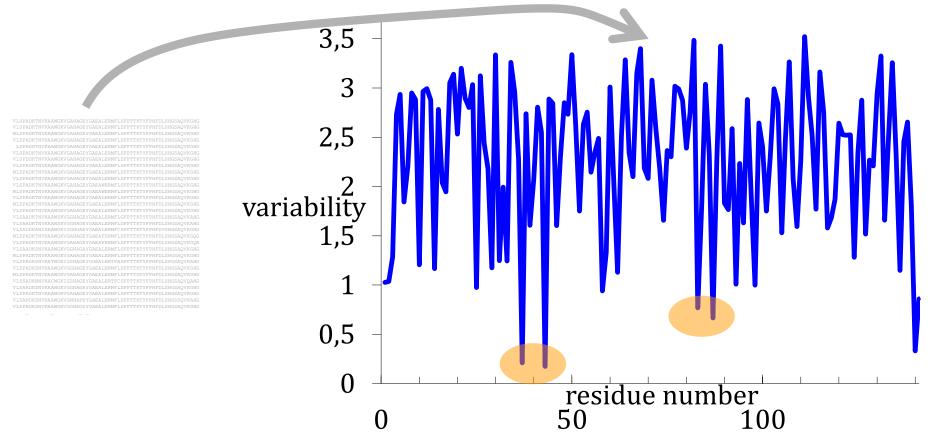
#### haemoglobin as example

summarise this data

VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG  $mostly \ for \ proteins^{_{\!\!\text{MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG}}_{^{\!\!\text{MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG}}$ VLSPADKTNVKAAWGKVGAHAGEYGAEALEKMFLSFPTTKTYFPHFDLSHGSAQVKGHG LSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKTNVKAAWGKVGAHAGDYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPDDKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKTHVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKTNVKAAWGKVGAHAGEYGAEAWERMFLSFPTTKTYFPHFDLSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEAWERMFLSFPTTKTYFPHFDLSHGSAOVKGHG VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAOVKGHG VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSAADKTNVKAAWSKVGGHAGEYGAEALERMFLGFPTTKTYFPHFDLSHGSAQVKAHG VLSAADKTNVKAAWSKVGGHAGEYGAEALERMFLGFPTTKTYFPHFDLSHGSAQVKAHG VLSADDKANIKAAWGKIGGHGAEYGAEALERMFCSFPTTKTYFPHFDVSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEAFERMFLSFPTTKTYFPHFDLSHGSAQVKGQG VLSPADKTNVKAAWGKVGAHAGEYGAEAFERMFLSFPTTKTYFPHFDLSHGSAQVKGQA VLSAADKSNVKAAWGKVGGNAGAYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKSNVKATWDKIGSHAGEYGGEALERTFASFPTTKTYFPHFDLSPGSAOVKAHG VLSPADKSNVKAAWGKVGGHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTGTYFPHFDLSHGSAQVKGHG VLSSADKNNVKACWGKIGSHAGEYGAEALERTFCSFPTTKTYFPHFDLSHGSAQVQAHG VLSAADKSNVKAAWGKVGGNAGAYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VLSANDKSNVKAAWGKVGNHAPEYGAEALERMFLSFPTTKTYFPHFDLSHGSSOVKAHG VLSPADKSNVKAAWGKVGGHAGDYGAEALERMFLSFPTTKTYFPHFDLSHGSAOVKGHG

#### **Conservation / variability**

#### Look at residues 37, 43, 83 and 87



- how do we get these and what does it mean?
- what does it mean for this protein ?

#### **Conserved residues**

Proximity to haem group

• green residues

• more on pro 37 later

# **Beliefs in multiple sequence alignments**

Similar proteins found in many organisms

- where they are conserved connected with function
- variation reflects evolution (phylogeny)

How many homologues might you have ?

- many
  - some DNA replication proteins almost every form of life
  - profilin cell mobility bacteria, mammals, plants
  - .
- few
  - exotic viral proteins
  - messengers exclusively in human biochemistry

# **Trees / Phylogeny**

Multiple sequence alignments are fun

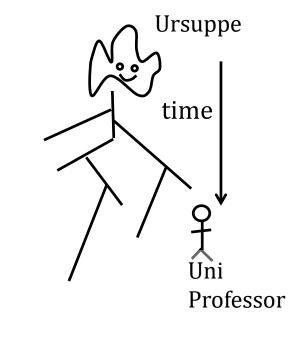
• conservation, function...

What next ? Phylogeny - making trees

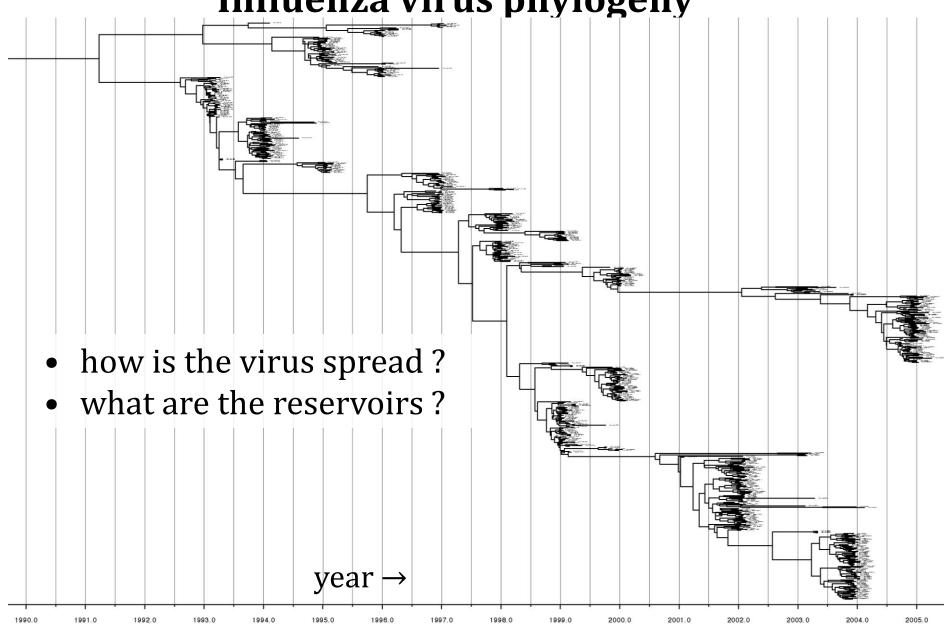
Need multiple sequence alignments to make trees

Do you just want the tree of life ?

- who killed the bananas ?
- where does influenza come from ?
- lassa, swine flu, ebola
- who killed the ladies ?

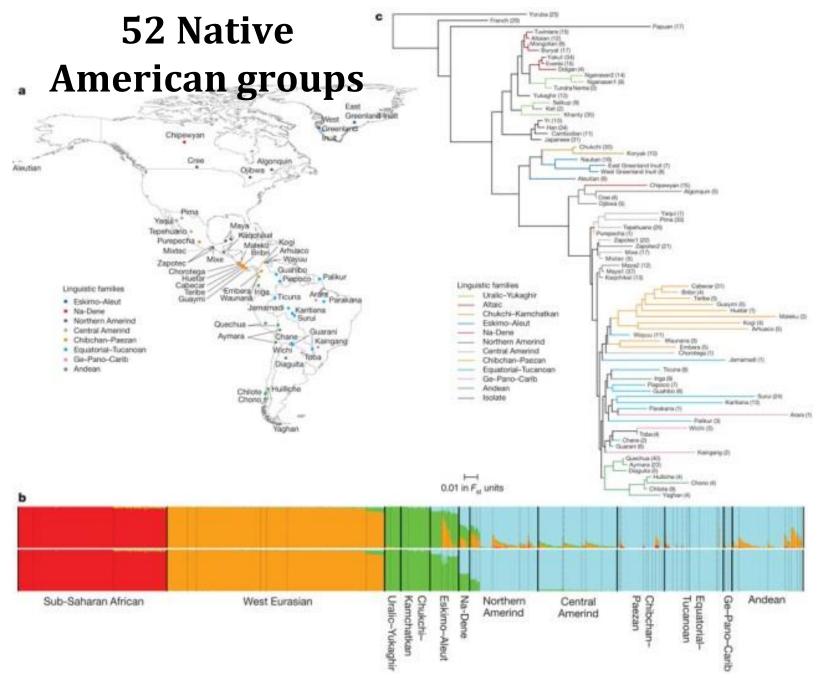


#### Influenza virus phylogeny



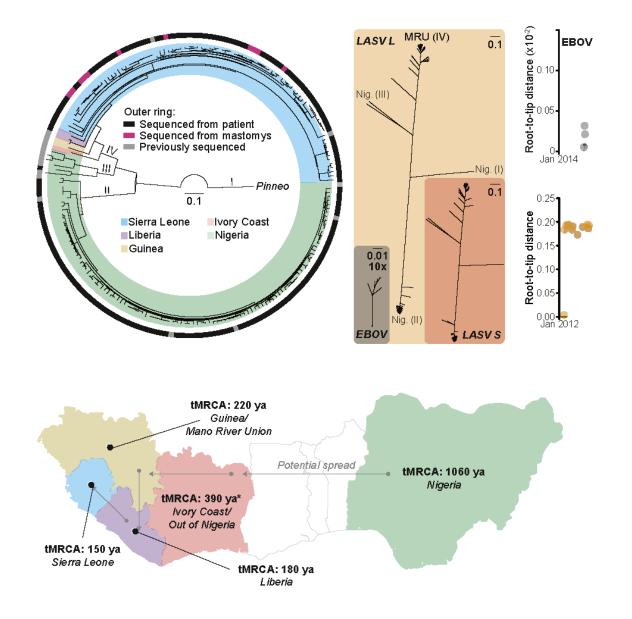
Rambaut, A., .. Holmes, C. The genomic.. influenza A virus, Nature 452, 1-6, 2008

29/06/2017 [7]



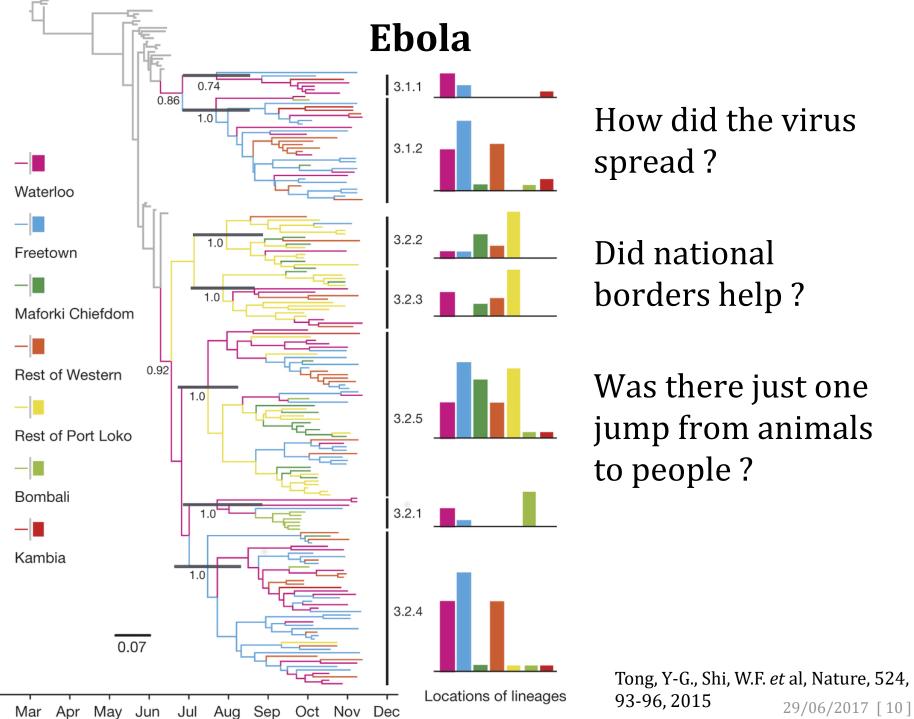
Reich, D., ...Ruiz-Linares, A., Nature, 488, 370 (2012), Reconstructing Native American .. History

#### lassa virus



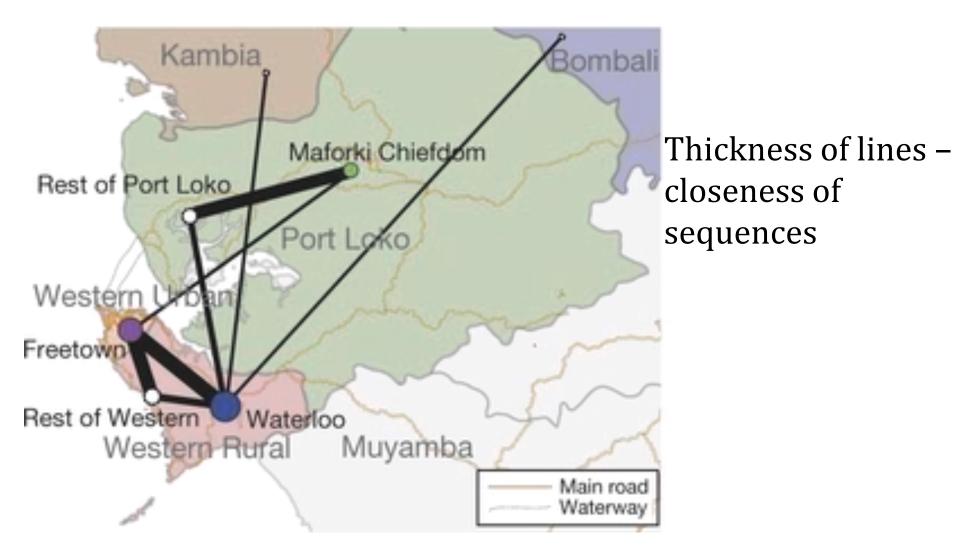
Andersen, KG... Sabeti, PC, Cell, 162, 738-750 (2015), "Clinical sequencing .. lassa virus"

29/06/2017 [9]



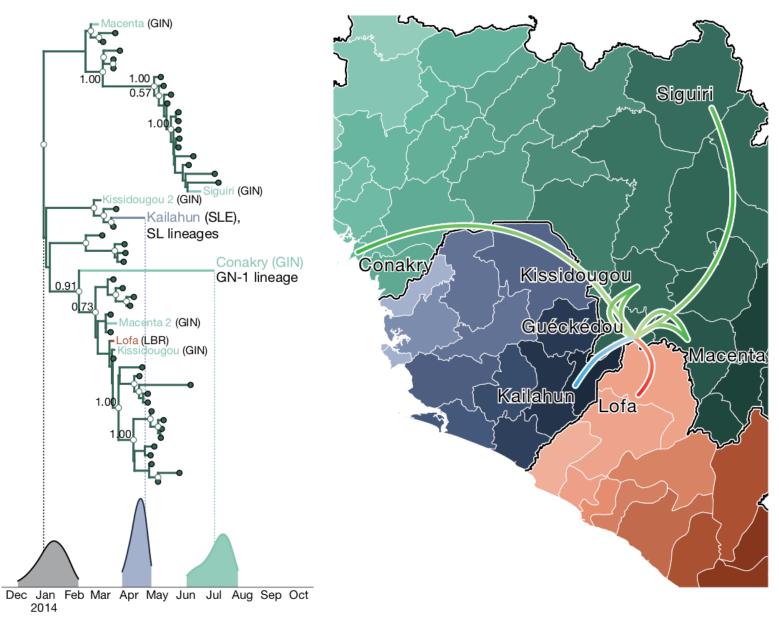
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#### How did the virus spread?

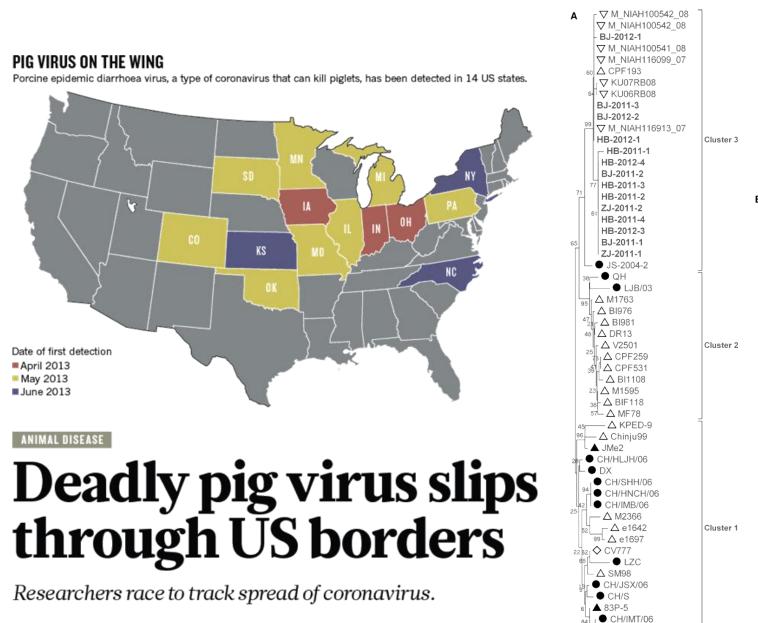


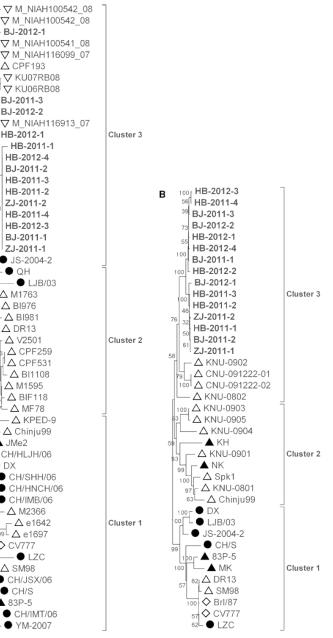
Tong, Y-G., Shi, W.F. *et* al, Nature, 524, 93-96, 2015

#### **Ebola and borders**



Dudas, G., ... Rambaut, A., Nature, 544, 309-315, 2017





0.005

Mole, B., Nature, 499, 388 (2013) Gao, Y. Kou, Q., Ge, X, Zhou, L., Guo, Yang, H., Arch Virol, 158, 711-715 (2012)

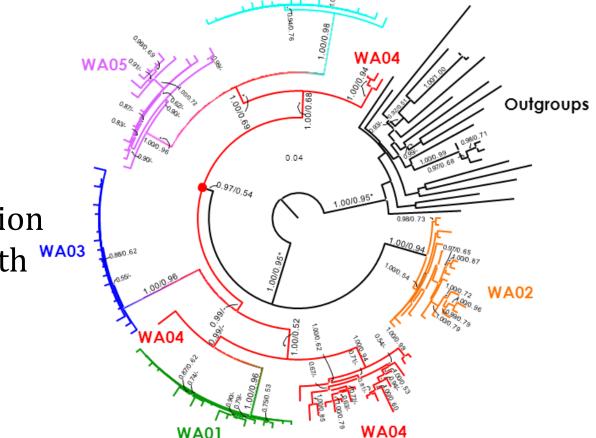
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# 1000 acts of sex

"the defendant intended to inflict "great bodily harm"... Between 1999 and 2004, he engaged in more than 1000 oral, vaginal, and anal acts of unprotected sex with his female partners"

black: normal population colours: associated with WA03 Herr WA04



Scaduto, D.I., Brown, JM, Haaland, WC, Zwicki, DJ, Hillis, DM, Metzker, ML, (2010) Proc Natl Acad Sci USA, 107, 21242 29/06/2017 [14]

# The plan

• optimise and alignment and tree simultaneously

#### Many sequences - rigorous alignment

- two sequence alignment
  - optimal path through  $n \times m$  matrix
- three sequence alignment
  - optimal path through  $n \times m \times p$  matrix
- four sequence alignment
  - ...
- *m* sequence alignment of *n* residues....  $O(n^m)$

Excuse to use lots of approximations

• no guarantee of perfect answer

Reasonable starting point

• begin with pairs of proteins

## **Scoring schemes**

$$S_{a,b} = \sum_{i=1}^{N_{res}} \operatorname{match}(s_{a,i}, s_{b,i})$$

In pairwise problem

VLSPADKSNVKAGWGQVGAHAGDYGAEAIERMYLSFPSTKTYFPHTDISHGSAQVKGHG MLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG

- Sum over match()
  N<sub>res</sub> is sequence length
- match(s<sub>a,i</sub>, s<sub>b,i</sub>) is the match/mismatch score of sequence a and b at position i
- invent a distance between two sequences like

$$d_{a,b} = \frac{1}{S_{a,b}}$$

distance measure..
 which sequences are most dissimilar to each other

# Scoring schemes for a multiple alignment

In the best alignment

- 1 is aligned to 2, 3, ..
- 2 to 3,4, ...

VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG
 VITP-EQSNVKAAWGKVGAHAGEYGAEALEQMFLSYPTTKTYFP-FDLSHGSAQIKGHG
 MLSPGDKTQVQAGFGRVGAHAG--GAEALDRMFLSFPTTKSFFPYFELTHGSAQVKGHG
 VLSPAEKTNIKAAWGKVGAHAGEYGAEALEKMF-SYPSTKTYFPHFDISHATAQ-KGHG
 -VTPGDKTNLQAGW-KIGAHAGEYGAEALDRMFLSFPTTK-YFPHYNLSHGSAQVKGHG
 VLSPAEKTNVKAAWGRVGAHAGDYGAEALERMFLSFPSTQTYFPHFDLS-GSAQVQAHA
 VLSPDDKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG

• then I move 5 and 2 & 5 and 3 – messes up 2 and 3

Mission: for  $N_{seq}$  sequences

•  $S_{a,b}$  : alignment score sequences a and b

$$score = \sum_{b \neq a}^{N_{seq}} \sum_{a=1}^{N_{seq}} S_{a,b}$$

- not quite possible
  - this method is just an approximation

# Aligning average sequences

VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VITPAEKTNVKAAWGKVGAHAGEYGAEALEQMFLSYPTTKTYFPHFDLSHGSAQIKGHG

#### and

IITPGDKTNVKAAFGKVGAHGGEYGAEALDRMFISFPSTKTYYPHFDLSHASAQVKAHG VITPAEQTNIKGAWGQIGAHAGDYAADALEQMFLSYPTSKTYFPYFDLTHGSAQIKGHG VITPAEKTQVKAAWGKVGGHAGEYGAEAIEQMFLTYPTTQTYFPHFELSHGTAQIKGHG

- At each position
- use some kind of average in scoring
- if a column has 2×D and 1×E score
  - score as 2/3 D + 1/3 E
- later.. call the average of S1 and S2: av(S1, S2)

# Summarise ingredients

- pairwise scores + distances
- ability to align little groups of sequences

# **Progressive alignments**

Guide tree / progressive / neighbour joining method

Steps

- build a distance matrix
- build a guide tree
- build up overall alignment in pieces

## **Progressive alignment - tree**

Compute pairwise

- S1 ATCTCGAGA
- S2 ATCCGAGA
- S3 ATGTCGACGA
- S4 ATGTCGACAGA
- S5 ATTCAACGA

alignments, S1 calculate the S2 .11 distance matrix S3 .20 .30 .27 S4.36 .09 .30 .33 S5 .23 .27 S1 S2 S3 S4 S5 S1 calculate guide tree S2 S3 S4 S5

## Multiple alignment from guide tree

- gaps at early stages remain
  Problems..
  - S1/S2 and S3/S4 good
  - no guarantee of S1/S4 or S2/S3

• av(S1,S2) is average of S1 and S2

#### align S1 with S2

S1	ATCTCGAGA
S2	ATC-CGAGA

align S3 with S4		
S3	ATGTCGAC-GA	
S4	ATGTCGACAGA	

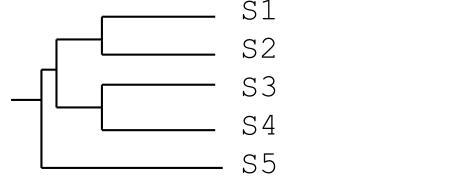
#### align av(S1,S2) with av(S3,S4)

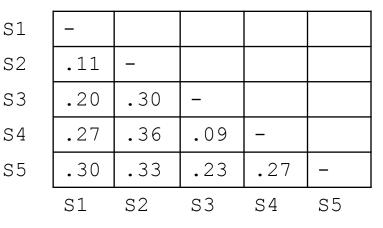
S1	ATCTCGAGA
S2	ATC-CGAGA
S3	ATGTCGAC-GA
S4	ATGTCGACAGA

#### align av(S1,S2,S3,S4) with S5

- S1 ATCTCGA--GA
- S2 ATC-CGA--GA
- S3 ATGTCGAC-GA
- S4 ATGTCGACAGA
- S5 AT-TCAAC-GA

#### **Problems and variations**

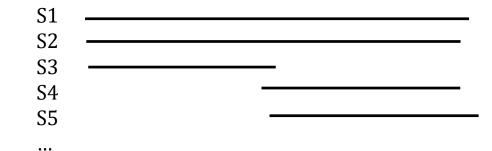




What order should we join ?

- pairs are easy (S1+S2) and (S3+S4)
- which next?

Real breakdown



S1 and S2 are multi-domain proteins

- S3 is not really related to S4 or S5
- distance matrix elements are rubbish

# Given an alignment

How reliable / believable ?

- set of very related proteins (an enzyme from 100 mammals)
  - no problem
- diverse proteins (an enzyme from bacteria to man)
  - lots of little errors
- can break completely (domain example)

Is the tree a "phylogeny"? A reflection of evolution?

• more later

#### **Measuring conservation / entropy**

Entropy

- how much disorder do I have ?  $S = -k_b \sum_{i=1}^{N_{states}} p_i \ln p_i$
- in how many states may I find the system?
- Our question
- look at a column how much disorder is there ?

VLSPADKTNVKAAWGKVC AHAGEYGAEALERMFLSFPTTKTYFPHE DI.SHGSAQVKGHG VITP-EQSNVKAAWGKVC AHAGEYGAEAIEQMFLSYPTTKTYFP-E DI.SHGSAQIKGHG MLSPGDKTQVQAGFGRVC AHAG--GAEAVDRMFLSFPTTKSFFPYE EI.THGSAQVKGHG VLSPAEKTNIKAAWGKVC AHAGEYGAEAAEKMF-SYPSTKTYFPHE DI.SHATAQ-KGHG -VTPGDKTNLQAGW-KIC AHAGEYGAEALDRMFLSFPTTK-YFPHY NI.SHGSAQVKGHG VLSPAEKTNVKAAWGRVC AHAGDYGAEAGERMFLSFPSTQTYFPHE DI.S-GSAQVQAHA VLSPDDKTNVKAAWGKVC AHAGEYGAEALERMFLSFPTTKTYFPHE DI.SHGSAQVKGHG

> no disorder

much disorder

Calculate an "entropy" for each column

# Entropy

- forget k<sub>b</sub> (Boltzmann just scaling)
  We have a protein
- 20 possible states.. use log base 20

$$S = -\sum_{i=1}^{N_{states}} p_i \log_{20} p_i$$

If a residue is always conserved ?  $p_i = 1$  or  $p_i = 0$ 

 $S = \log_{20} 1 = 0$  (no entropy)

What if all residues are equally likely ?  $p_i = \frac{1}{20}$ 

$$S = -\sum_{i=1}^{20} \frac{1}{20} \log_{20} \frac{1}{20} = -20 \cdot \frac{1}{20} \log_{20} \frac{1}{20} = -20 \cdot \frac{1}{20} (-1)$$

= 1

• my toy alignment...

# Entropy

- First column is boring

#### - Second

$$p_{\rm D} = \frac{5}{7}$$
  
 $p_{\rm E} = \frac{1}{7}$ 

 $p_{\rm N}={}^1\!/_7$ 

VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDISHGSAQVKGHG VITP-EQSNVKAAWGKVGAHAGEYGAEAIEQMFLSYPTTKTYFP-FDISHGSAQIKGHG MLSPGDKTQVQAGFGRVGAHAG--GAEAVDRMFLSFPTTKSFFPYFEITHGSAQVKGHG VLSPAEKTNIKAAWGKVGAHAGEYGAEAAEKMF-SYPSTKTYFPHFDISHATAQ-KGHG -VTPGDKTNLQAGW-KIGAHAGEYGAEALDRMFLSFPTTK-YFPHYNISHGSAQVKGHG VLSPAEKTNVKAAWGRVGAHAGDYGAEAGERMFLSFPSTQTYFPHFDIS-GSAQVQAHA VLSPDDKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDISHGSAQVKGHG

$$S = -\left(\frac{5}{7}\log_{20}\frac{5}{7} + \frac{1}{7}\log_{20}\frac{1}{7} + \frac{1}{7}\log_{20}\frac{1}{7}\right)$$
  
\$\approx 0.27\$

#### **Entropy from DNA**

Exactly as for proteins (use  $p_i \log_4 p_i$ )

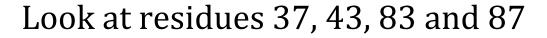
max possible entropy

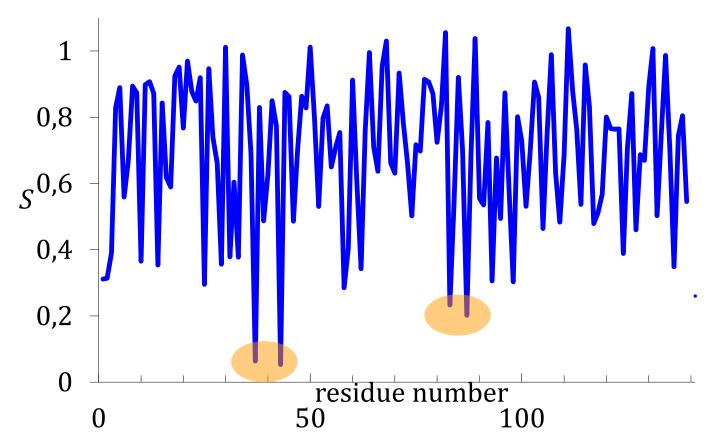
$$S = -4\left(\frac{1}{4}\log_4\frac{1}{4}\right)$$
$$= -4\left(\frac{1}{4}\cdot(-1)\right)$$

= 1

example from start of this topic

#### Haemoglobin conservation





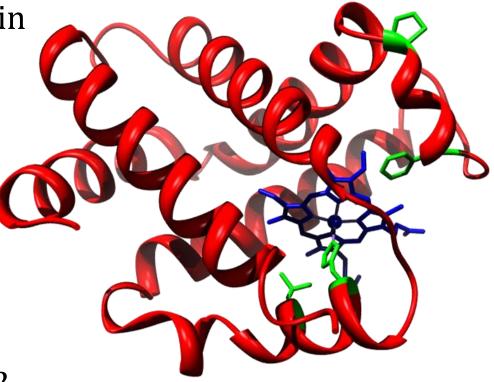
4 residues (maybe more) stand out as conserved

• why ?

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# **Conserved residues in haemoglobin**

- 3 of the sites are easy to explain
- interact with haem group
- Look at fourth site
- proline
- end of a helix



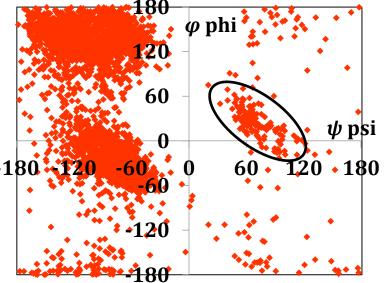
What is special about proline?

- no H-bond donor
- here if it mutates, maybe haemoglobin does not fold

#### **Conservation for structure**

Some residues have very special structural roles

- proline not an H-bond donor
  - often end of a helix
- glycine can visit part of  $\varphi \psi$  plot
  - found in some turns



Are all gly residues so important?

• NO – they occur in many places sometimes in turns

Are all pro residues very conserved ? No

#### **Conservation for function**

In a serine protease

- always a "catalytic serine"
- can it mutate ? Not often

In haemoglobin – residues necessary for binding haem

- can they mutate ? rarely
- changes properties of haemoglobin (bad news)

Dogma

residues in active site will be more conserved than other sites

#### **Important summary**

Conservation may reflect

- important function
- structural role
- Mutagenesis / chemistry
- what residue may I change to allow binding to a solid substrate ? (for biosensor/immobilized enzyme ?)
- try error prone PCR to select for new enzyme activity which sites might I start with (active site) ?
- Drug design example
- target is an essential protein (basic metabolism, DNA synthesis, protein synthesis..)
- is there some set of sequence features common to pathogen, different to mammalian protein ?

#### **Evolution – do not trust conservation**

Imagine: two possible systems for some important enzyme

- 1. active site fits to essential biochemistry

  - any mutation you lose active site residues are conserved in a conservation plot
- 2. maybe enzyme is not absolutely perfect
  - some mutations kill you
  - some mutations OK lacksquare
  - site does not appear perfectly conserved •

Where would you evolve to?

- 1. very fragile
- 2. likely to survive mutations

Resistance to mutations...

#### **Tolerance of mutations**

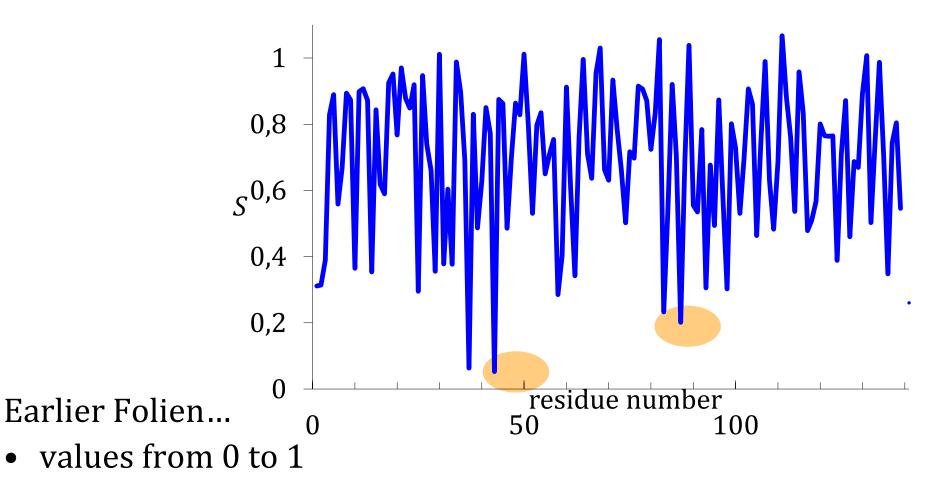
Boring answer

• some amino acids are similar to each other

Better answer – if your protein can tolerate mutations

- your genes have a better chance of being passed on
- will be selected for
- it is a Darwinian trait

#### **Conservation – how meaningful ?**



What if I used more homologues?

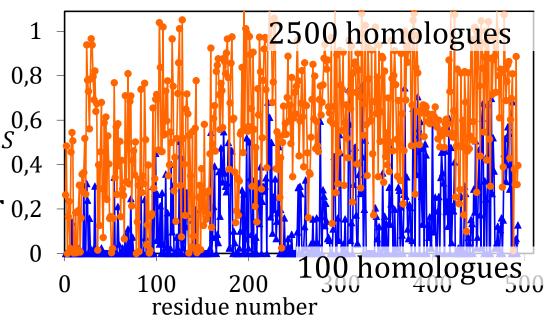
### **Conservation – how meaningful ?**

Example sequence (DNA gyrase)

- find 100 close homologues (mostly > 80% similarity)
   calculate conservation
- find 2500 close homologues (mostly > 50 % similarity) calculate conservation

Fewer sequences

- lots of conserved sites
- you can get the answer 0,2 you want 0



Consequences - summarise

### Significance of conservation

You read in a paper – residue 37 is conserved

- how many sequences did they look at ?
  - 10 ? bad, 100 better, 1000 better
- choosing the number of sequences lets you manipulate results
- statistically
  - have you sampled enough sequences ?

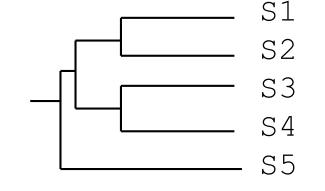
## **Phylogeny / Evolution**

The trees in text books are almost never perfect One rarely knows the correct history

Problems..

Previously we had a "guide tree"

did (S1,S2) and (S3,S4) share an ancestor but not S5 ?



- branch lengths do not reflect evolutionary time
- there may be other similar trees which could be evolutionary paths

### **Evolutionary time**

Compare two DNA sequences see 1 mutation (represents time *t*) 2 mutations (time 2*t*) 3 mutations (time 3*t*)... No !

#### After some evolution

 $A \rightarrow C \rightarrow G$ two events (although looks like  $A \rightarrow G$ ) $A \rightarrow C \rightarrow G \rightarrow C \rightarrow A$ looks like zero mutations

If I have infinite time

- all bases / residues equally likely
- $p_{mut} = 3/4 = 0.75$  (DNA) or  $p_{mut} = 19/20$  (protein)

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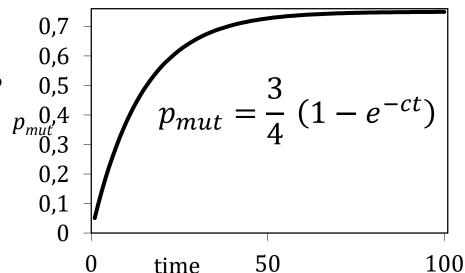
## **Mutation probability**

- time units are arbitrary
- how would I estimate time ? (for DNA)

• 
$$t \propto -\ln\left(1-\frac{4}{3}p_{mut}\right)$$

•  $p_{mut}$  ? count  $\frac{n_{mut}}{n_{res}}$ 



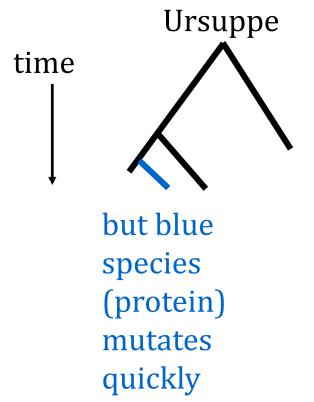


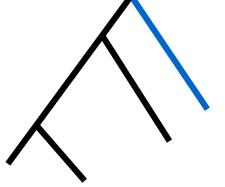
For short times,  $p_{mut}$  changes fast

- for small *t*, distances will be more reliable
  - as will be alignments
- Is this enough for phylogeny ?
- what about reliability ?

## **Problems in phylogeny**

- not all sites mutate equally quickly
- not all species mutate equally quickly





blue appears to have branched off earlier

- Backwards mutations?
- not really a problem (Klausurerfahrung)

### **Problems estimating time**

- 1. mutation rates vary wildly
  - changing environments pH, temperature,...
- 2. imagine time *t* is such that  $p_{mut}$ = 0.25
  - we have random events
  - sometimes you see 23% mutation, sometimes 28%
- time estimates will never be accurate
- maybe we cannot find the correct tree
  - can we roughly estimate reliability?

## Reliability

# Think of first alignment

VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHFDLSHGSAQVKGHG VITP-EQSNVKAAWGKVGAHAGEYGAEAIEQMFLSYPTTKTYFP-FDLSHGSAQIKGHG MLSPGDKTQVQAGFGRVGAHAG--GAEAVDRMFLSFPTTKSFFPYFELTHGSAQVKGHG VLSPAEKTNIKAAWGKVGAHAGEYGAEAAEKMF-SYPSTKTYFPHFDISHATAQ-KGHG -VTPGDKTNLQAGW-KIGAHAGEYGAEALDRMFLSFPTTK-YFPHYNLSHGSAQVKGHG VLSPAEKTNVKAAWGRVGAHAGDYGAEAGERMFLSFPSTQTYFPHFDLS-GSAQVQAHA

### What would happen if you deleted a column?

- if the data is robust /reliable
  - not much
- if the tree is very fragile /sensitive
  - tree will change

better...

## Reliability

Repeat  $10^2$  to  $10^3$  times

- delete 5 to 10 % of columns
- copy random columns so as to have original size
- recalculate tree

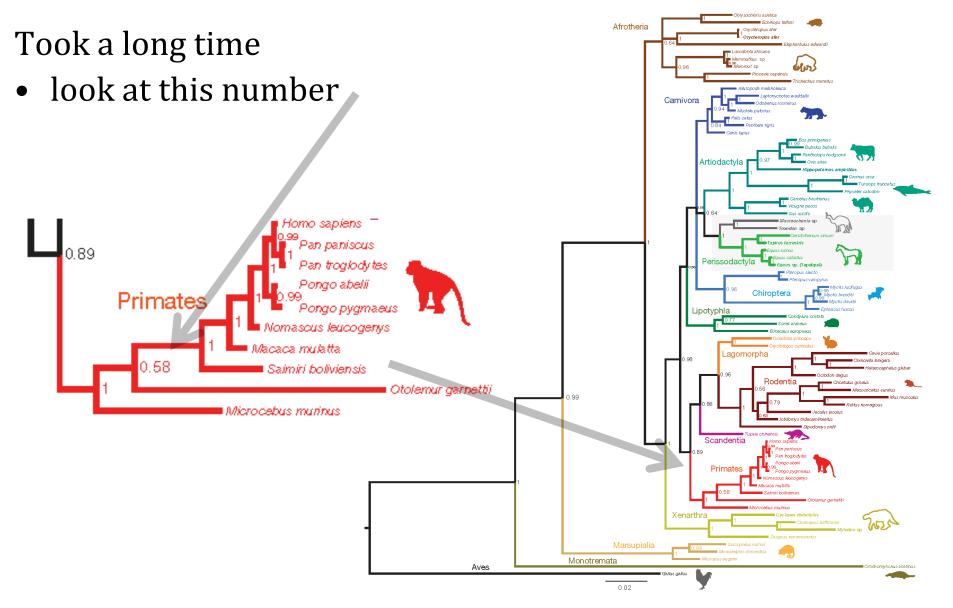
How often did you see each branch?



### Monster example

- generate 1000 trees
- for each sub-tree
  - see how often it is present
- example from nature

### **Monster calculation**



Welker, F., ... MacPhee, D.E. Nature, 522, 81-84 (2015) Ancient proteins resolve the evolutionary ...

29/06/2017 [48]

### **DNA or protein sequences ?**

The issues

- regulatory regions, RNA genes
- synonymous mutations (common only seen in DNA)
- non-synonymous mutations (amino acid changes)
  - more information  $D \rightleftharpoons E$ ,  $I \rightleftharpoons L \rightleftharpoons V$ , ...

Alignment reliability

- proteins
  - uses codon structure (implicitly)
  - better, amino acid similarity,  $I \rightleftharpoons L \rightleftharpoons V$  is not bad
- DNA
  - less information

## **DNA or protein sequences ?**

	protein	DNA	time
synonymous changes	not seen	yes	short
a.a. changes	yes	yes	longer
a.a. similarity	accounted for	not seen	
frame shifts	not seen	yes	
non-coding regions	not helpful	yes	

Very short time or not protein-coding

• use DNA

Longer time and coding for protein

• use proteins

## Summary

- multiple sequence alignment conservation
  - find important residues (function or structure)
  - can quantify conservation
- relations between most similar proteins are most reliable
- best tree is never found
  - too difficult algorithmically
  - lots of errors evolution is a random process
- rough idea of reliability
- quick tree possible for 1000s of sequences
- more complicated methods phylogeny in Biologie courses