

Analysis, Comparison of Proteins

Andrew Torda, wintersemester 2009 / 2010, GST

From previous lectures

- we know about protein structures / coordinates
- we know how coordinates are collected

What kind of analysis would we like to do ?

- recognising common features
- classifying
 - (useful for prediction)

Philosophy

- ways to measure similarity between structures
- ways to find similar pieces / "motif"s
- most common motif ? Secondary structures

Next few weeks

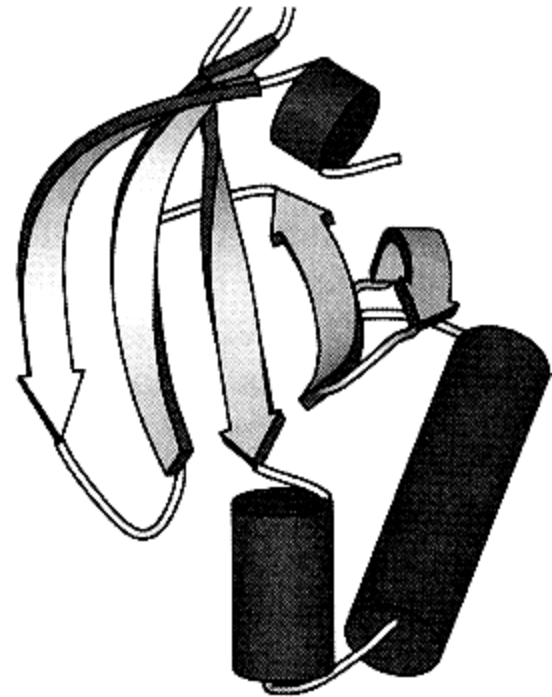
- Secondary structure definitions
- Classifying protein structures
- Domains
- Supersecondary structure
- Protein similarity – sequence versus structure
- Sequence space
- Classifications – hierarchical
- Classifications – other
- Comparison of proteins

- touching on evolution, alignments, ...

Secondary Structure Recognition



from coordinates



assumes structures
recognised

- how to define / recognise secondary structure ?

Defining Secondary Structure

- What do I want ?

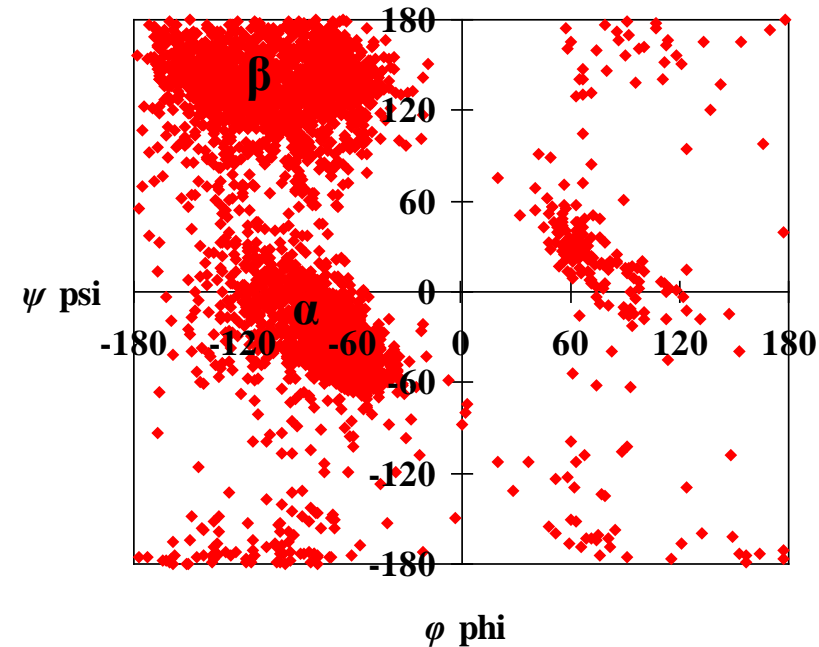
	1	2	3	4	5	6	7	8	9	10	...
residue	A	C	A	D	L	V	A	W	W	A	...
sec struct	-	H	H	H	H	H	-	E	E	E	...

- at each residue, label as to secondary structure type
 - no ambiguity
 - labels at residues – not between !
- I do not want probabilistic answers (more soon)
- remember not all residues are in recognisable α -helix or β -sheet

Secondary Structure From Coordinates

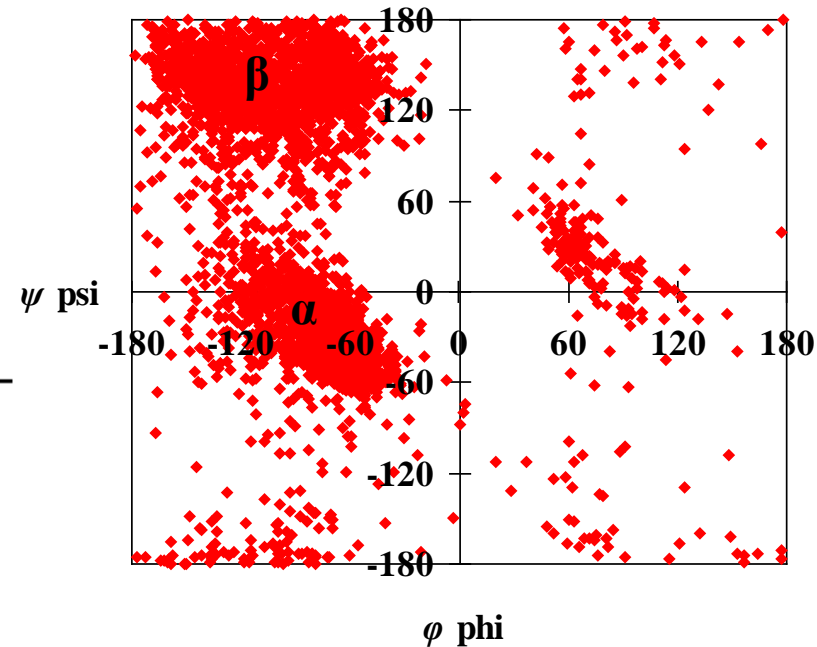
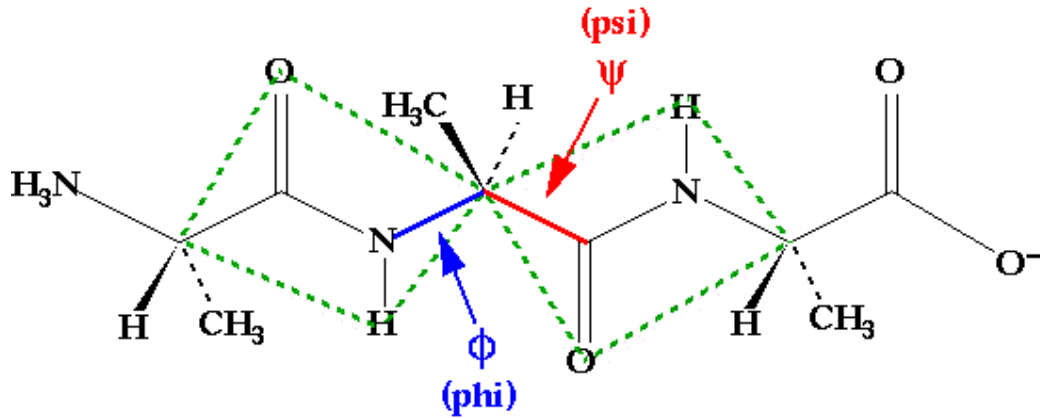
Start with α -helices

- what do we know
 - look like helices
 - 3.6 residues per turn
 - H-bond pattern
 - N residue i to $i+4$
- residue backbone angles



Using Backbone Angles

Given coordinates, easy to calculate

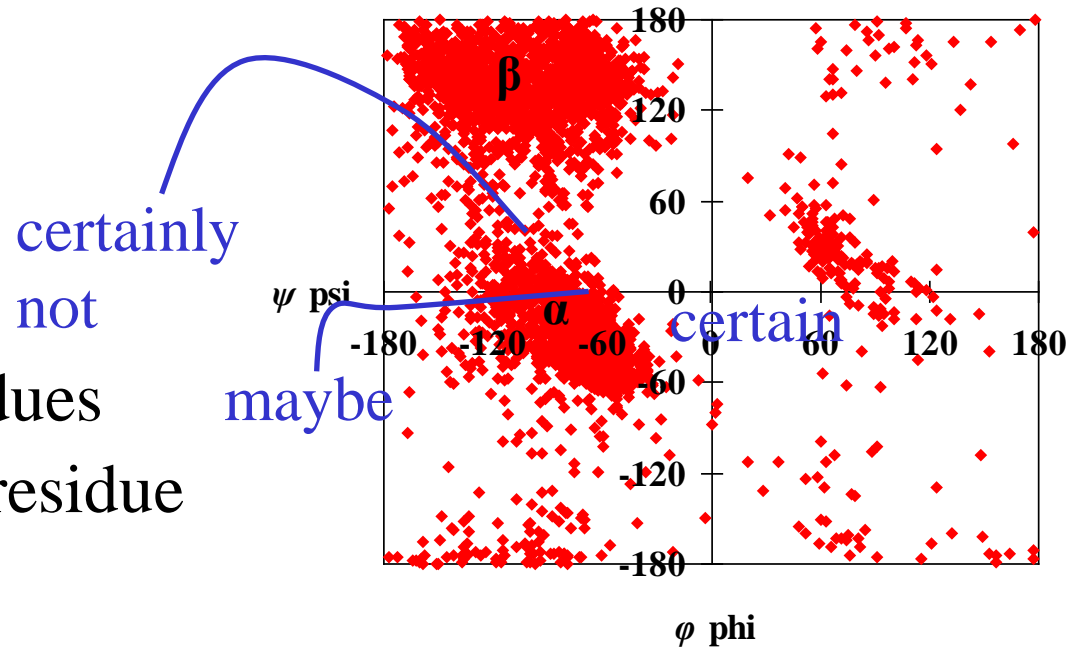


Problems

- what are my thresholds ?
- what if I see one residue with angle ?

Problems With Using Angles

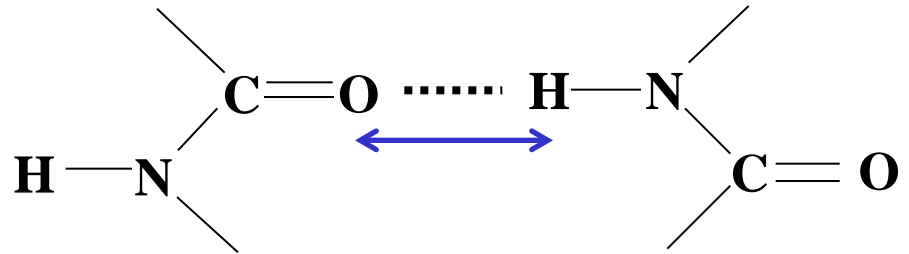
- thresholds
- minimum number of residues
 - what if I see only one residue with perfect angles ?
 - not forming H bonds
 - need 3 or 4 residues



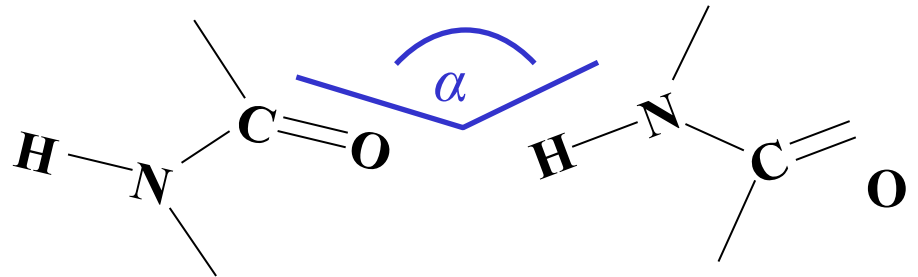
Maybe We Should Use H-Bonds

We have the coordinates

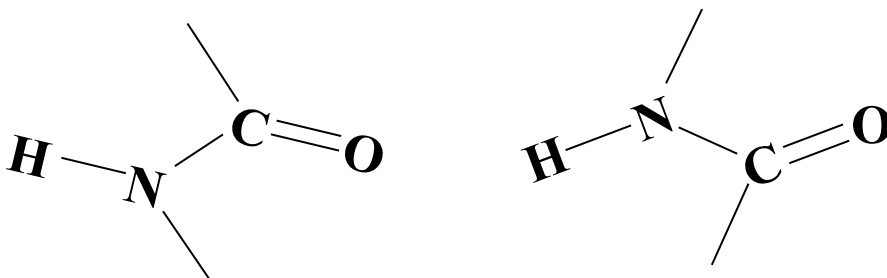
- should be easy to recognise all H bonds
- criteria ?
- distance $r(\text{ON}) < \approx 3.6 \text{ \AA}$



- angle ? ($\approx 120^\circ$)



A practical definition



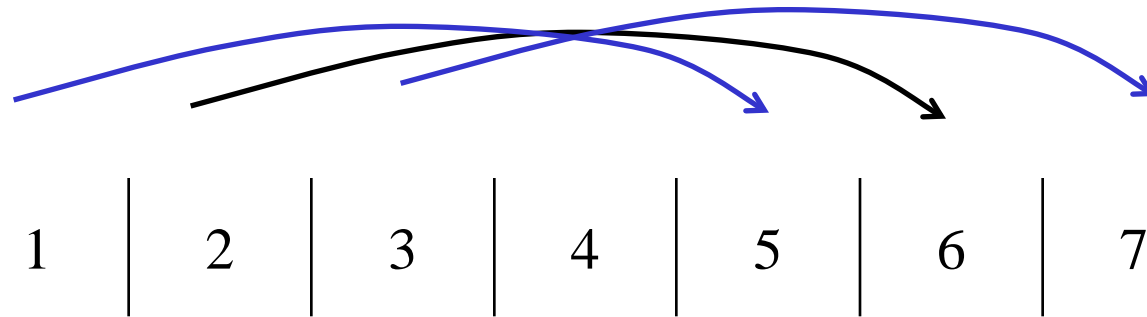
$$E = 332 \, q_1 q_2 \left(\frac{1}{r_{\text{ON}}} + \frac{1}{r_{\text{CH}}} - \frac{1}{r_{\text{OH}}} - \frac{1}{r_{\text{CN}}} \right)$$

- require $E < -3$ (arbitrary !)
- note as r grows, E goes to 0

Problems with short helices

What if I see only 3 or 4 residues ?

- real helix has 2 H-bonds per residue
- what if I see one ?



Compromise

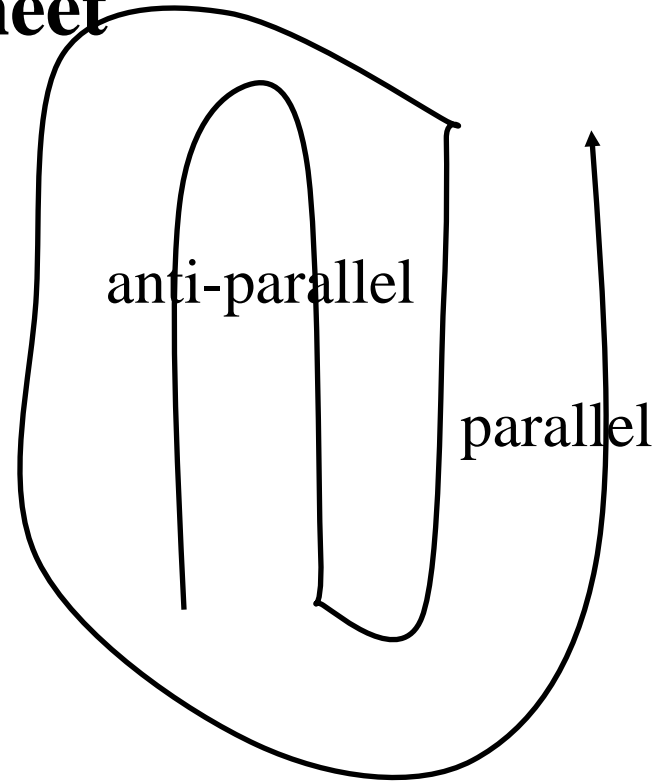
- call this a turn (only has one H bond)
- α -helix definition
 - at least two consecutive (4 residue) turns

Useful definitions (α -helix)

- recognise an H-bond $i, i+4$ (either
 - $r_{\text{ON}} + \text{angle } \alpha$ or
 - general distance formula $= \text{turn}$
- two successive turns
 $= \text{minimal } \alpha\text{-helix}$
- more overlapping helices
 $= \text{longer helix}$
- all we have done is an α -helix

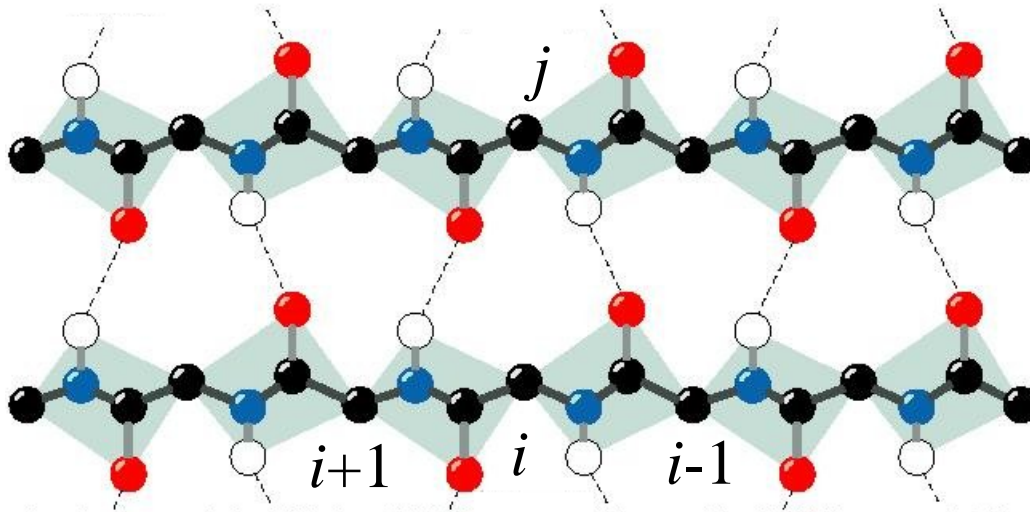
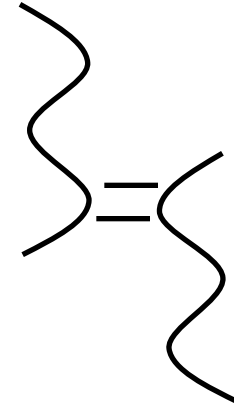
a β -strand / sheet

- much more difficult
 - parallel versus anti-parallel
 - H-bond neighbours not known
 - 5-109
 - 6-110
 - 7-111 ... parallel
- or
 - 5-109
 - 6-108
 - 7-107...anti-parallel
- formalise this



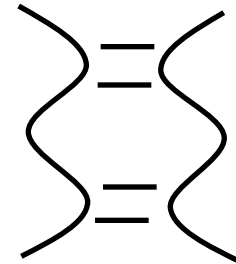
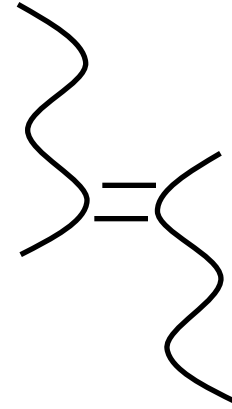
Defining a β -sheet

- start with a bridge
- parallel bridge
 - H-bond ($i-1, j$) & H-bond ($j, i+1$) or
 - H-bond ($j-1, i$) & H-bond ($i, j+1$)



Defining a β -sheet

- start with a bridge
- parallel bridge
 - H-bond ($i-1, j$) & H-bond ($j, i+1$) or
 - H-bond ($j-1, i$) & H-bond ($i, j+1$)
- ladder = one or more consecutive bridges
- sheet = one or more consecutive ladders with shared residues
- similar definition for anti-parallel sheets



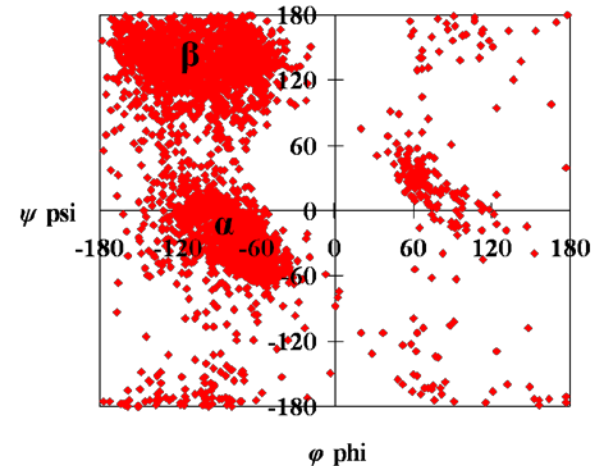
Are these problems real ?

Do thresholds matter ?

- do programs give the same answer ?
- if we use secondary structure for comparisons of proteins
- comparisons with experiment

Can we set perfect thresholds ?

- not all H-bonds are the same
- look at ψ - ϕ map, borders are not clear
 - mobility, finite energy
- coordinates have experimental error
- our programs generate worse coordinates (holes, distortions)



From secondary to higher levels Classification

- Why classify proteins ?
- Why recognise similarities
 - function prediction
 - structure prediction
 - vague idea of structure for mutagenesis, applications
- Why might this be useful ?
 - how many structures are there ?

How Many Protein Structures Are There ?

- Protein Databank $\approx 5.5 \times 10^4$
- 90 % sequence similarity $\approx 1.7 \times 10^4$
- different shapes 2 to 5×10^3
- implications for structure prediction ?
 - how many possible structures can we think of ?
 - exponential
 - how big is the real search space ?
 - really 10^3 to 10^4

Why So Few Structures

- discretization of space (makes it look smaller)
- physical reasons
 - compactness, stability
 - advantages of H-bonded conformations
- history / evolution
 - imagine all proteins evolve from some original molecule
 - evidence
 - theoretical – geometric constructions
 - chemical – construction of artificial protein(s)

Before Classifying

- earlier description of structure
- primary (sequence)
- secondary (α -helices, β -sheets, ...)
 - supersecondary ?
- tertiary
 - arrangement of helices / sheets or
 - where atoms are in space
- quaternary...
- we need idea of domains, then supersecondary structures

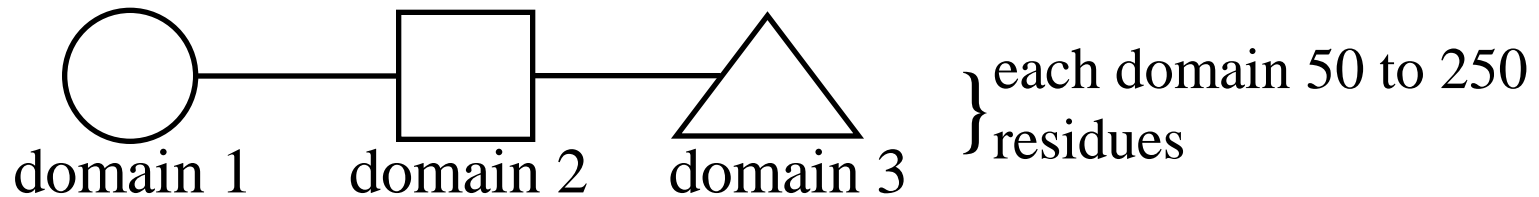
Domains in Biochemistry

History / biochemistry / no structures

- invented story
- we have a big protein
 - catalyses $A \rightarrow B$
 - C regulates it
- cleave protein (break with enzyme) to two parts
 - 1 still converts $A \rightarrow B$
 - 2 binds C
 - interpretation
 - catalytic domain
 - C binding domain
- more generally
 - different pieces of protein, responsible for different functions

Domain Concept Useful ?

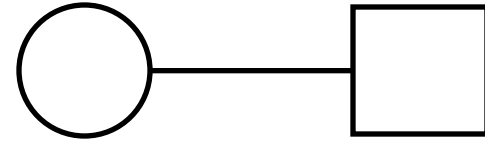
- Many times a whole protein cannot be crystallised, solved by NMR
- attack protein with enzymes to break up
- look for activity in pieces, solve structures of pieces



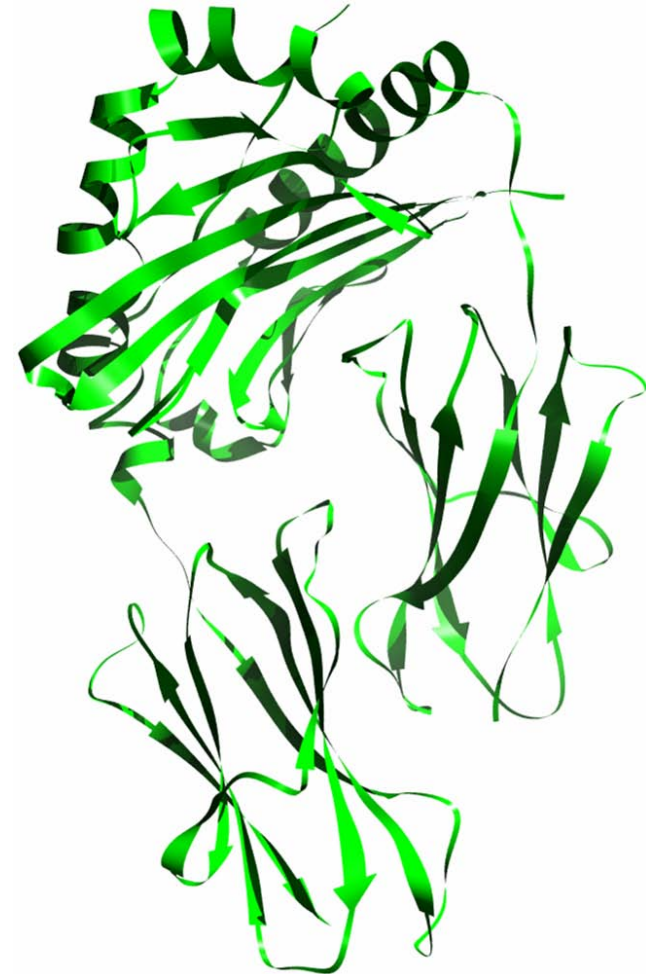
- literature / PDB full of "xxx domain of yyy"
- attractive ?
 - makes big proteins seem manageable
 - building block concept
 - attractive in evolutionary terms

Domains in Structures

- Many structures solved look like...



- histocompatibility module (1iak)
 - 3 domains + another protein
- are they always so clear ?
- porphobilinogen deaminase (1gtk)



Domain definition version 3

Three reasonable definitions

- biochemistry
- structures
- look for conserved units in sequence comparisons

Domains for today

- compact structural units

Domains for classification

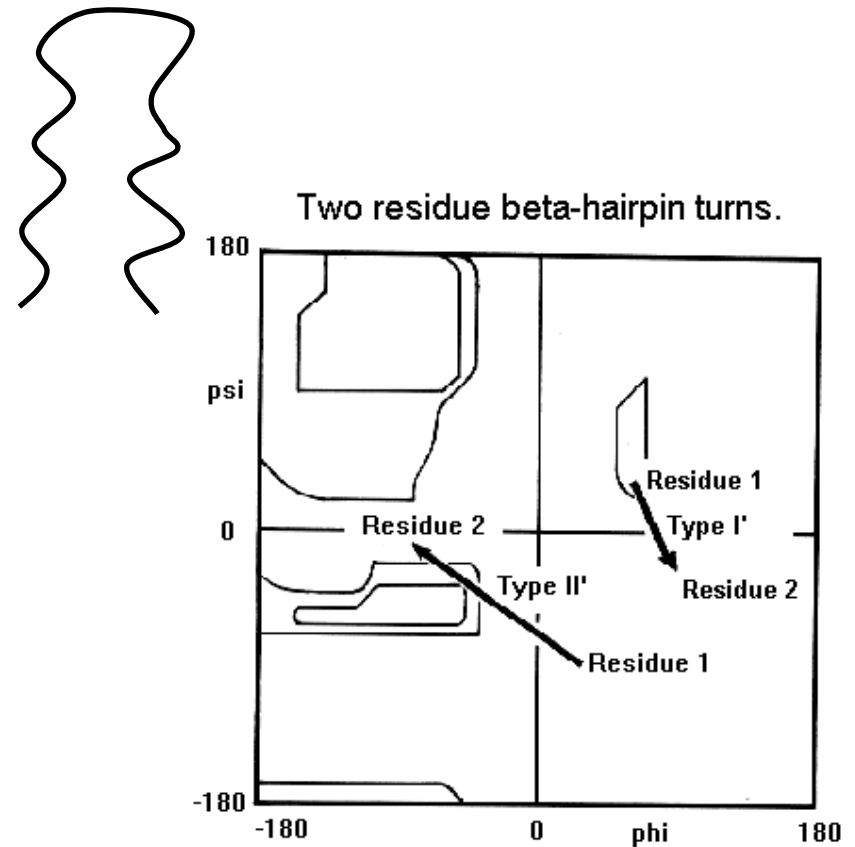
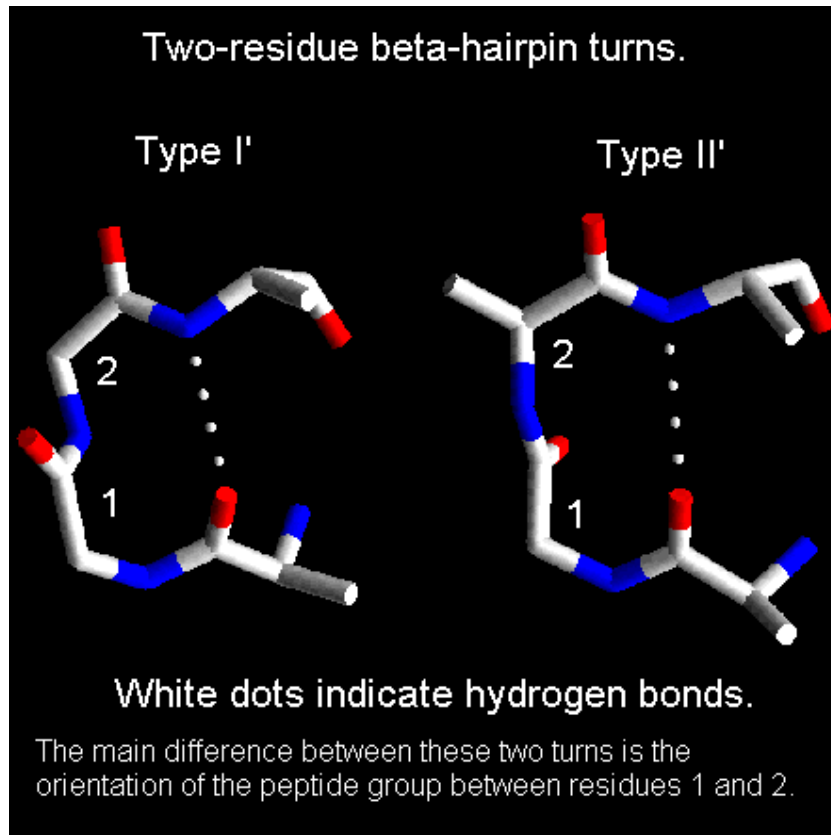
- structural classifications often domain based

Classifications In General

- 1. secondary structure
 - we see collections of residues and classify into recognisable types
- 2. different types of domain
 - soon
- 1b. supersecondary elements ?
 - are there some common small arrangements of α -helices, β -sheets ?

A Supersecondary Structure

- β -hairpin (β -turn- β) fits idea of common motif
 - described as built on secondary structure + specific H-bonds



More Supersecondary Structures

- helix-turn-helix (α -X- α)
 - DNA binding proteins
- helix-longer_loop-helix
 - Ca^{++} binding
- ...

Who cares ?

- repeated patterns / motifs suggests there are smaller number of structural units to recognise
- modularity appeals
- functional association
- conforms to some ideas on protein folding (more next semester)

Why I Do Not Like Supersecondary Structure

- ideal picture...
 - primary structure arrangement →
 - secondary structure / arrangement →
 - supersecondary structure →
 - tertiary structure or domains
- implies supersecondary structure is useful hierarchical element
 - not really used !

Sequence vs Structural Similarity

Background

- in the real world we usually have sequence information first
- want to make guesses about protein structure

I have two aligned protein sequences

- are they structurally similar ?

Old rule

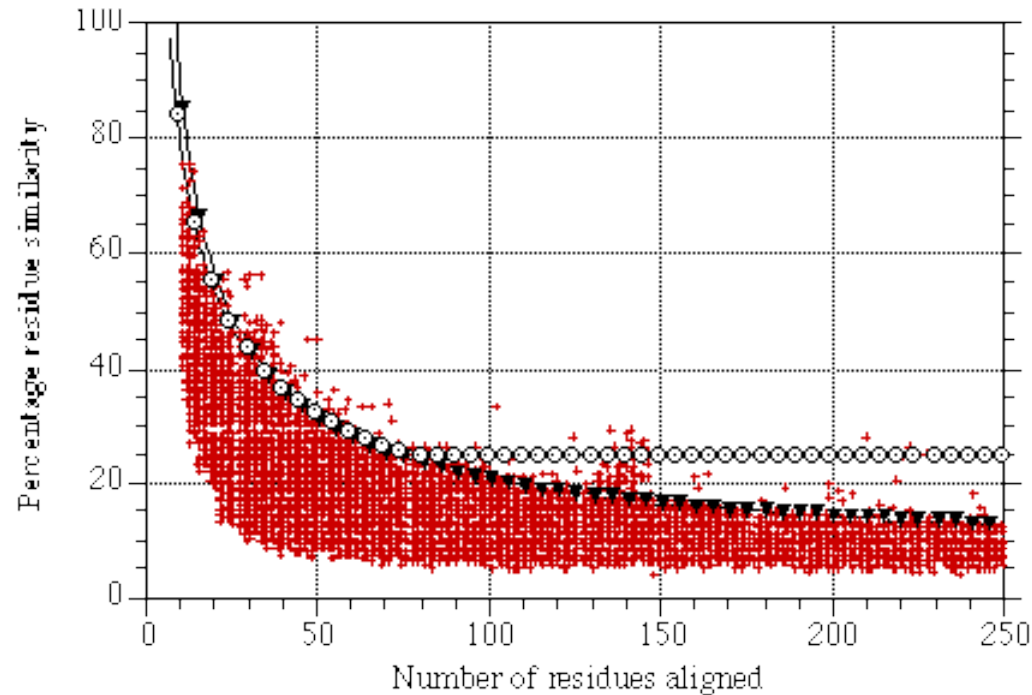
- $> 25\%$ sequence similar – similar structures
- $< 20\%$ cannot tell
- $20\% < x < 25\%$ - "twilight zone"

Is this universally valid ?

Sequence Similarity → Structure

Take a set of pairs of proteins

- find those which are not structurally similar
- look at sequence similarity
- old rule is not valid
- 50 residues
 - > 30 % seq
- 150 residues
 - > 20 %
- rule:
 - sequence similarity (length dependent) very good indicator of structural similarity



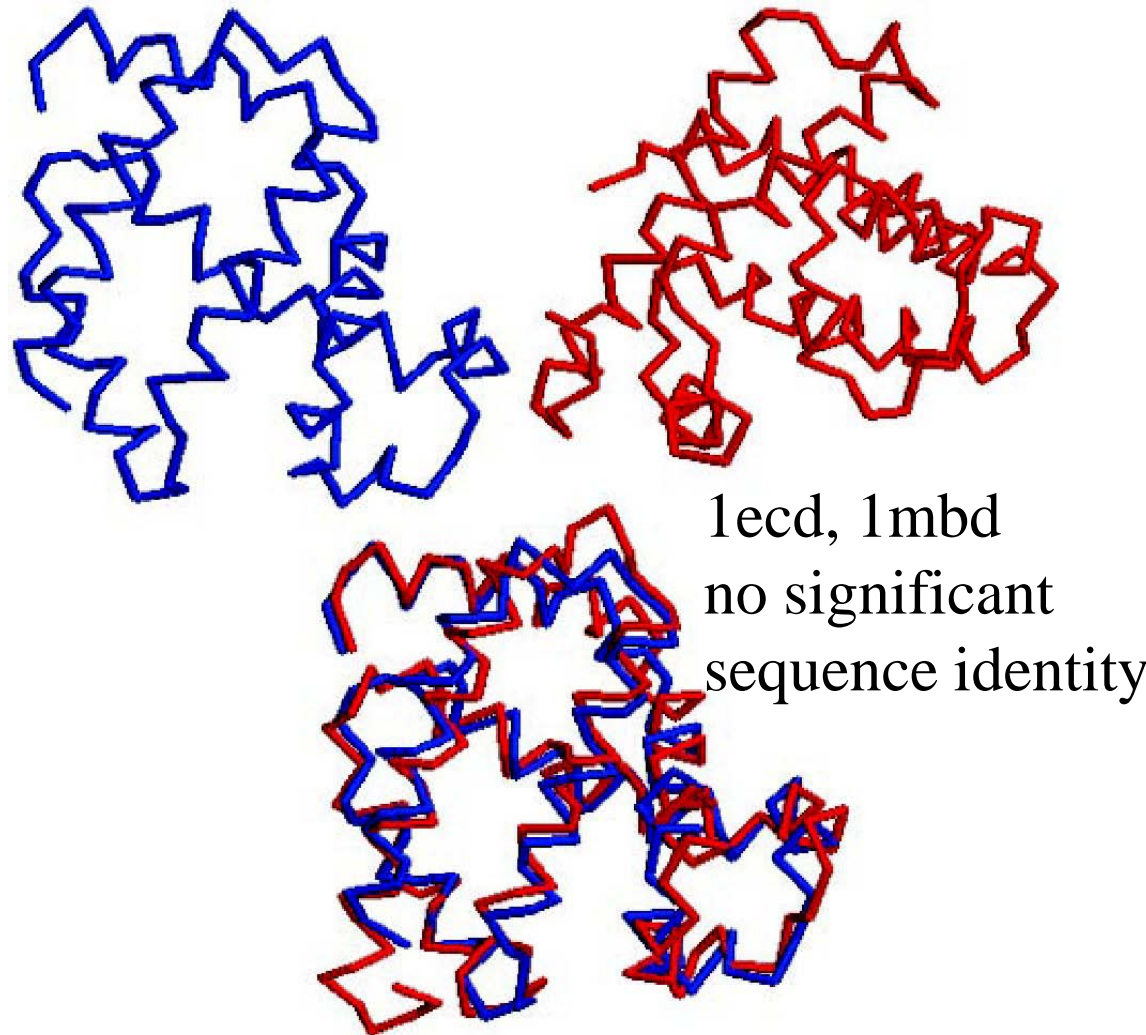
Using Sequence Similarity

- consequence
 - I could try to categorise proteins based on sequence
- tools
 - any alignment program (blast, fasta, clustal, ...)
- method
 - survey all proteins in the protein databank
 - collect all pairs $> x \%$ (or use more sophisticated threshold)
- result (jan 2009)

similarity	num clusters
90 %	20 002
70%	17 490
50%	14 906
- much more than 2 to 5×10^3 ?
- maybe some of my classes are not really different

Sequences Not Similar

- Sequences similar ? .. similar structure
- Sequences different ?
 - ??
- Example
 - 100's examples



An Example Family

- example, neighbours of 1cun chain A
 - look at sequence identity (%id)
 - alignment length (lali = number of residues)
 - root mean square diff in Å

No	Chain	%id	lali	rmsd	Description
1	1cunA	100	213	0.0	ALPHA SPECTRIN
2	1hcia	24	111	1.6	ALPHA-ACTININ 2
3	1ek8A	12	106	4.4	RIBOSOME RECYCLING FACTOR
4	1oxzA	9	91	2.5	ADP-RIBOSYLATION FACTOR BINDING PROTEIN GGA1
5	1ehlA	8	102	4.6	RIBOSOME RECYCLING FACTOR
6	1hx1B	5	105	3.1	HEAT SHOCK COGNATE 71 KDA
7	1dd5A	8	103	4.7	RIBOSOME RECYCLING FACTOR
8	1lvfA	9	98	2.6	SYNTAXIN 6
9	1bg1A	9	99	2.3	STAT3B
10	1hg5A	5	98	3.0	CLATHRIN ASSEMBLY PROTEIN SHORT FORM
11	1hs7A	14	92	2.5	SYNTAXIN VAM3
12	1dn1B	10	101	2.7	SYNTAXIN BINDING PROTEIN 1
13	1ge9A	6	108	4.6	RIBOSOME RECYCLING FACTOR
14	1fewA	8	125	3.5	SECOND MITOCHONDRIA-DERIVED ACTIVATOR OF
15	1qsdA	4	90	2.4	BETA-TUBULIN BINDING POST-CHAPERONIN COFACTOR
16	1e2aA	6	95	2.8	ENZYME IIA
17	1iliP	7	95	3.3	NEUROLYSIN
18	1fioA	8	100	2.6	SSO1 PROTEIN
19	1m62A	8	81	2.8	BAG-FAMILY MOLECULAR CHAPERONE REGULATOR-4
20	1k4tA	6	147	25.8	DNA T

DIVERSION Sequence Space

- convenient way to explain ideas of sequence similarity
- conventional spaces
 - 1D (x), 2D (x, y), 3D (x, y, z), 4D (x, y, z, w), ...
 - let us estimate how big a space or problem is
 - how many variables do I have ? (a, b, c, ...)
 - how many values can each variable have ?
 - a 3 values, b 4 values, c 5
 - number of points in space = $3 \times 4 \times 5$
- protein sequences
 - each position can have 1 of 20 values
 - total number of sequences = $20 \times 20 \times \dots = 20^{N_{res}}$
 - like a space of N_{res} dimensions

Representing a Sequence

- protein sequence and structural coordinates

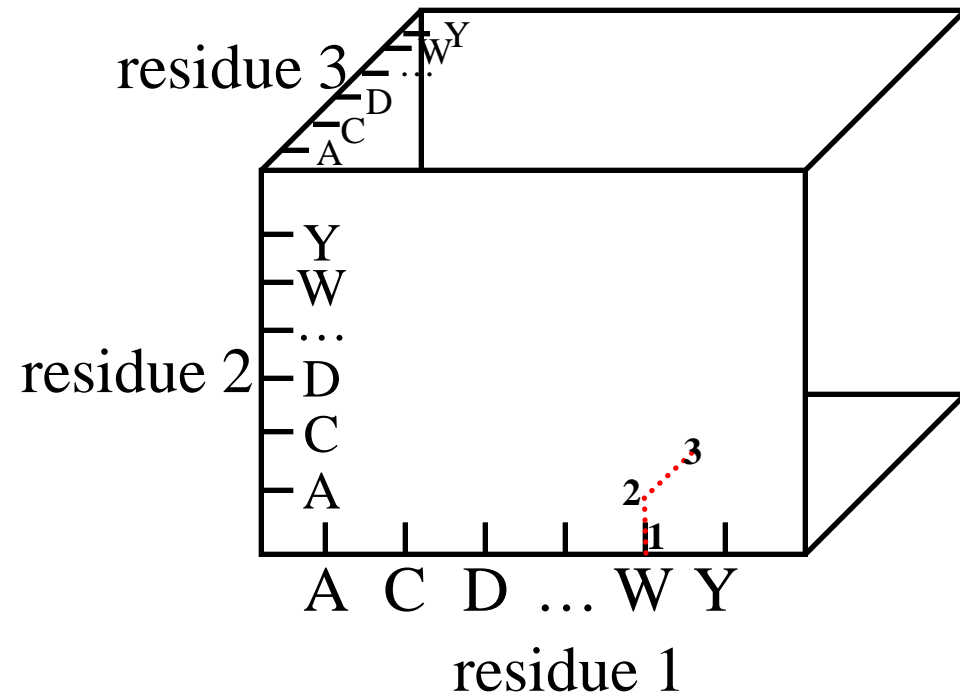
	1	2	3	4	5	6	7	...	N_{res}
x	1.2	2.3	...						10.3
y	2.4	3.5	...						11.1
z	1.7	2.9	...						15.5
seq	W	A	C	A	A	...			D

- consider the first three residues
 - WAC (for pictures only)

Finding a Sequence in This Space

- real diagram is a box of N_{res} dimensions
 - this one 3 dimensions

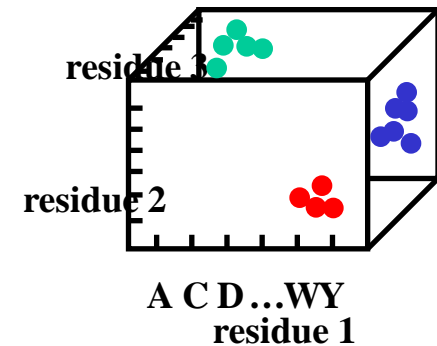
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seq	W	A	C	A	A	...			D



- looking for sequences...

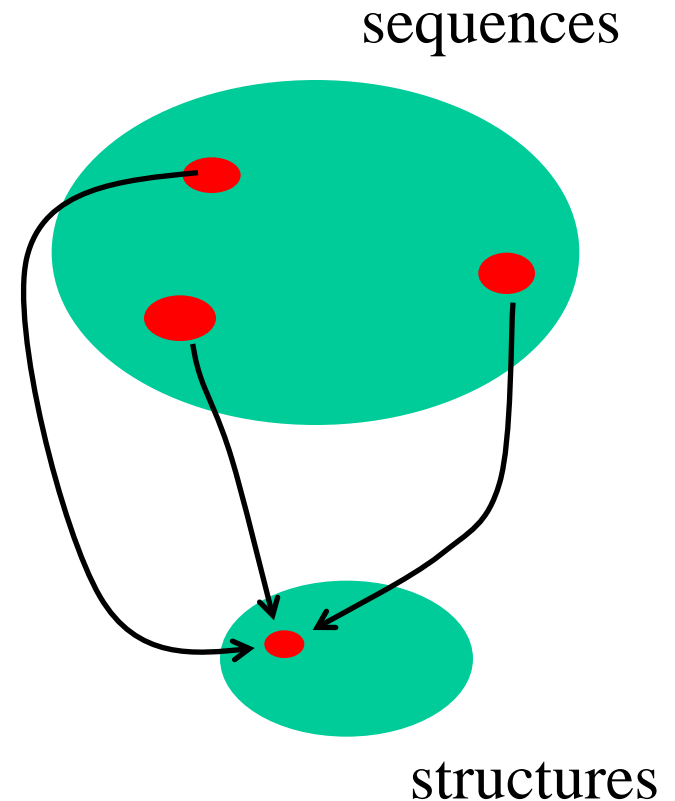
Families in Sequence Space

- Similar sequences should land near each other
- How realistic ?
 - picture is a simplification
 - only works for $N_{seq1} = N_{seq2}$
 - very useful
 - distances between sequences
- Will return next semester



Structure vs Sequence

- there are 1000's of such families
- summarise
 - similar sequences
 - similar structures
 - very different sequences
 - similar or different structures
- why ?



Structures < Sequences... Why ?

Evolution 1

- many small changes
- if structure changes, function breaks, you die
- sequences change as much as possible within this constraint

Evolution 2

- maybe some cases of convergent evolution
- impossible to prove

Consequences of sequence based categorisation

- we will have different classes, but really same protein shape

Surprising ?

- consider near universal proteins
 - 100's millions years evolution, function largely preserved
- chemistry
 - sequence does determine structure, many sequences could fit structure

Back to Classification

Sequence classification

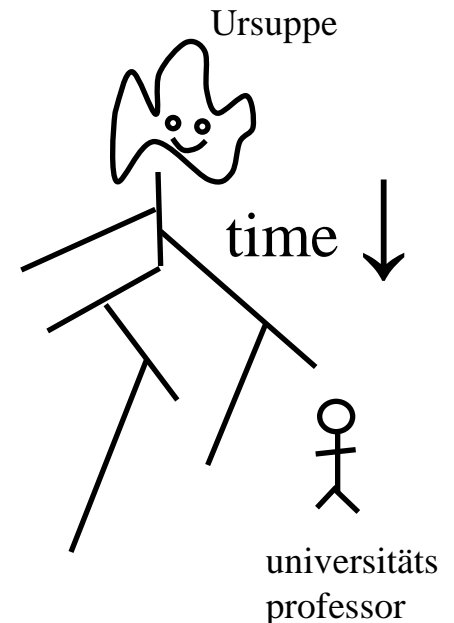
- good, reliable, similar class = similar structure
- not enough to find all similarities
- need for structure based methods

Philosophies

1. evolution
2. just classify proteins

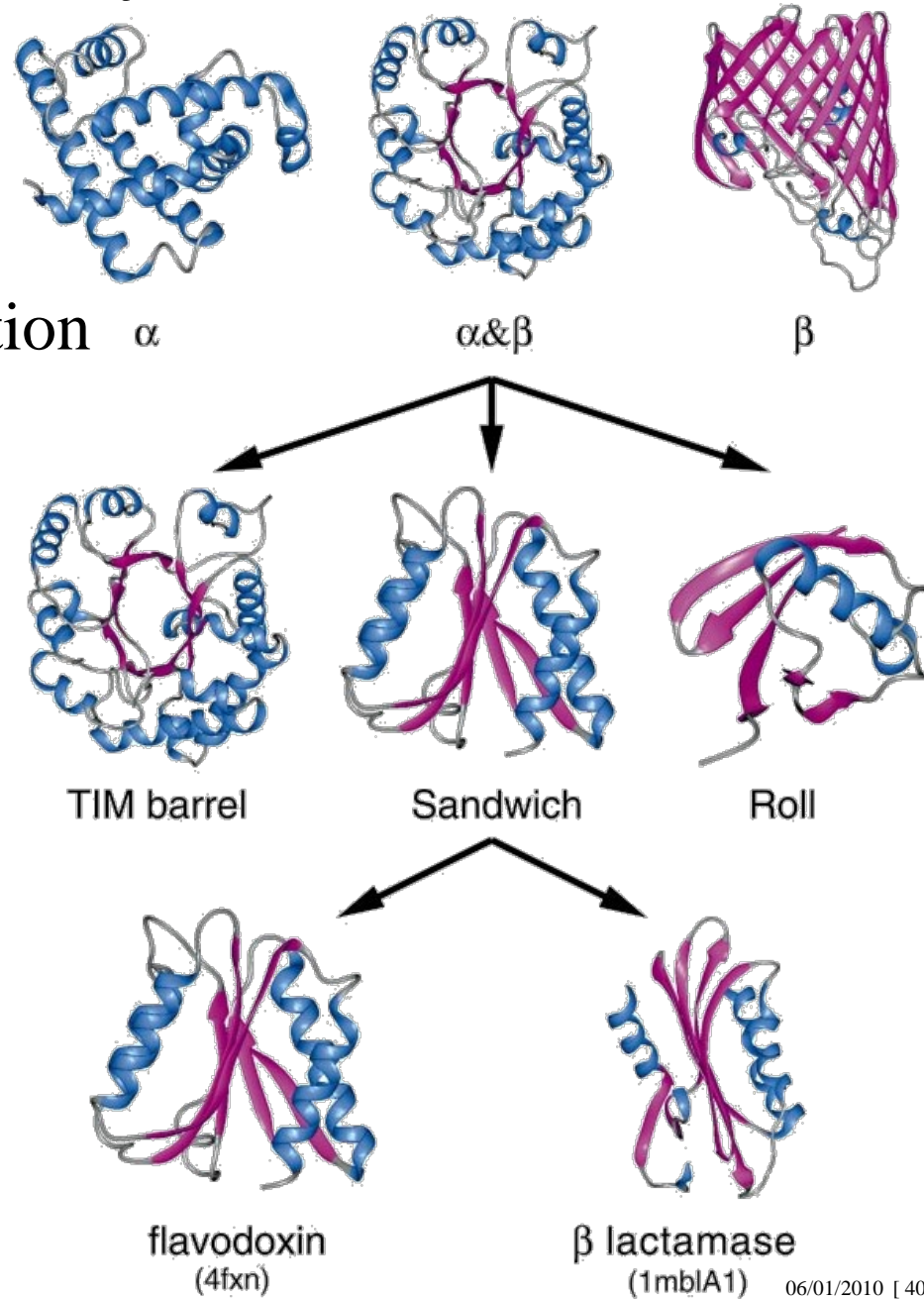
Evolution

- diagram →
- we expect a hierarchy

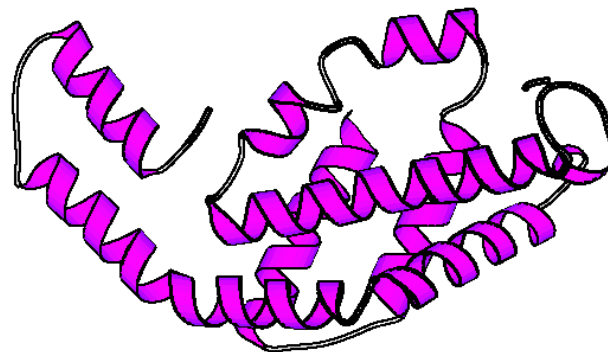
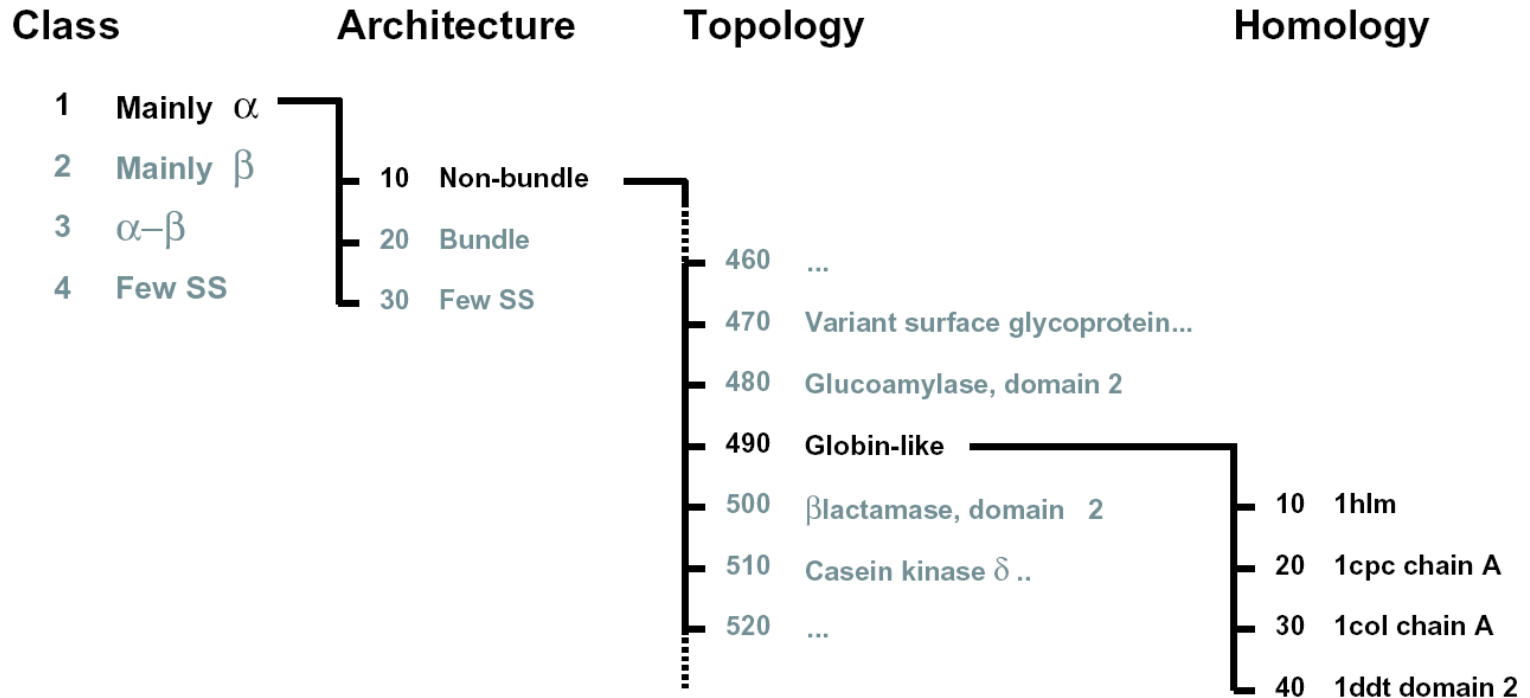


Imposing a Hierarchy on Proteins

- parts may correspond to evolution
- top level ?
- How useful and applicable ?
 - examples



Example from "CATH"



1.10.490.20

Mainly α .Non-bundle.Globin-like.1cpc chain A

Lots of families

α -helix bundles ?

- ≈ 226 domains, 3 % surveyed structures

β -sandwich ≈ 1236 domains, 15 %

some families ?

- < 0.01 %

Interesting...

- some families very popular, some not



α Bundle (2ccy)



α Non-Bundle (1eca)



α Few SS (2lfo)



β Ribbon (1pm)



β Single Sheet (1hre)



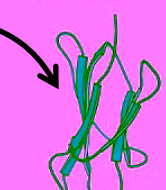
β Roll (1ph)



β Barrel (2por)



β Clam (3bc1)



β Sandwich (2h1aB)



β Distorted Sandwich (1edp)



β Trefoil (1afcA)



β Orthogonal Prism (1msaA)



β Altered Prism (1ymoA)



β 4 Propeller (1hxn)



β 6 Propeller (1nscA)



β 7 Propeller (2bbkH)



β 8 Propeller (2aahA)



β 2 Solenoid (1tsp)

Why are some families populated more than others ?

- more next semester
 - are some structures more stable ?
 - are some older in evolutionary terms ?
 - can some "accommodate" more sequences / tolerate more mutations
-
- is this a reflection of physics ?
 - no – PDB is very biased
 - mainly soluble, globular proteins which crystallised
 - very few membrane-bound proteins

Supersecondary Structure

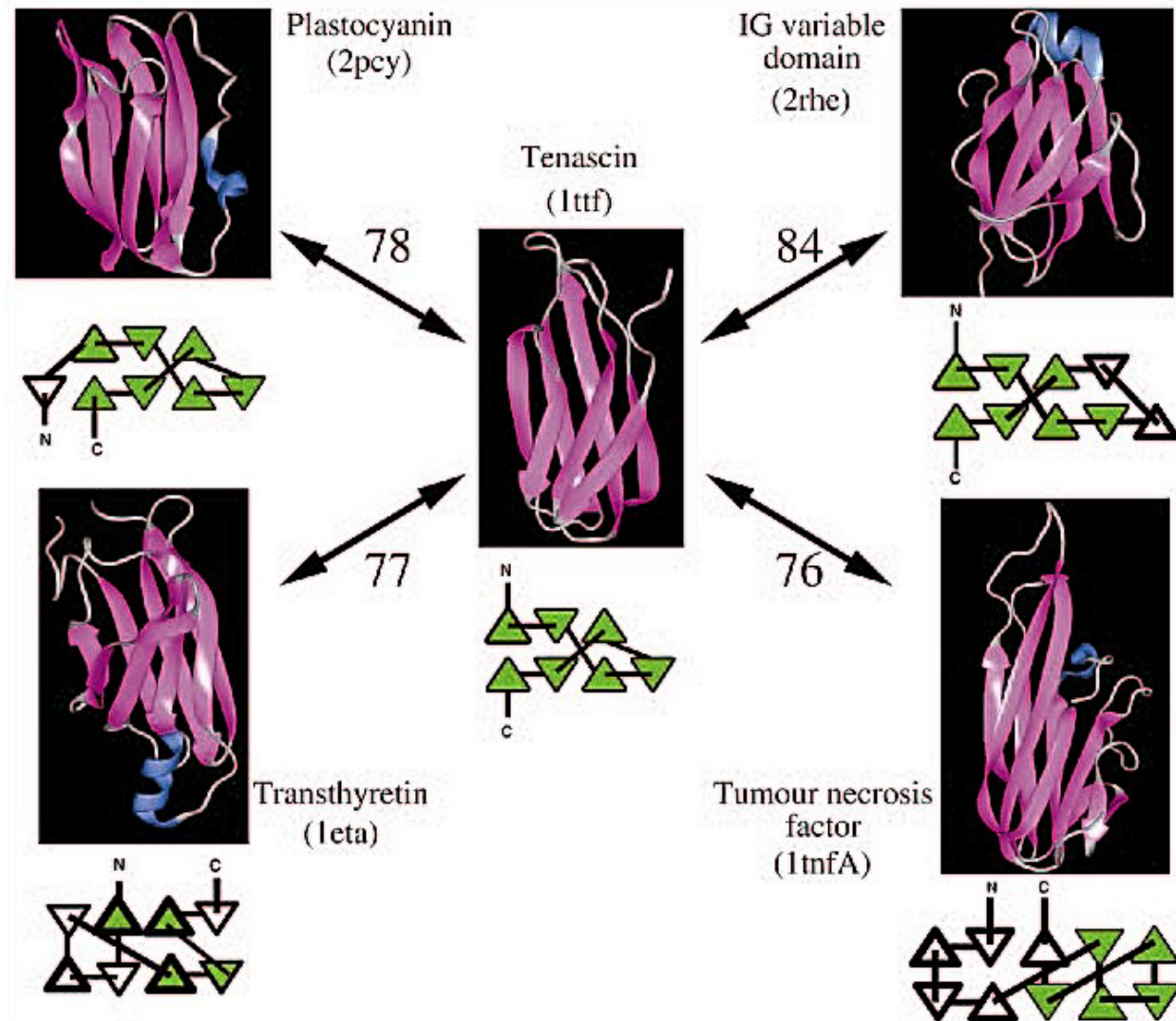
- Was supersecondary structure helpful here ?
- members of a given family probably have common supersecondary motifs.
 - not all proteins can be generated as a collection of motifs

Evolutionary interpretation

- given a classification – does it reflect evolution ?
 - maybe

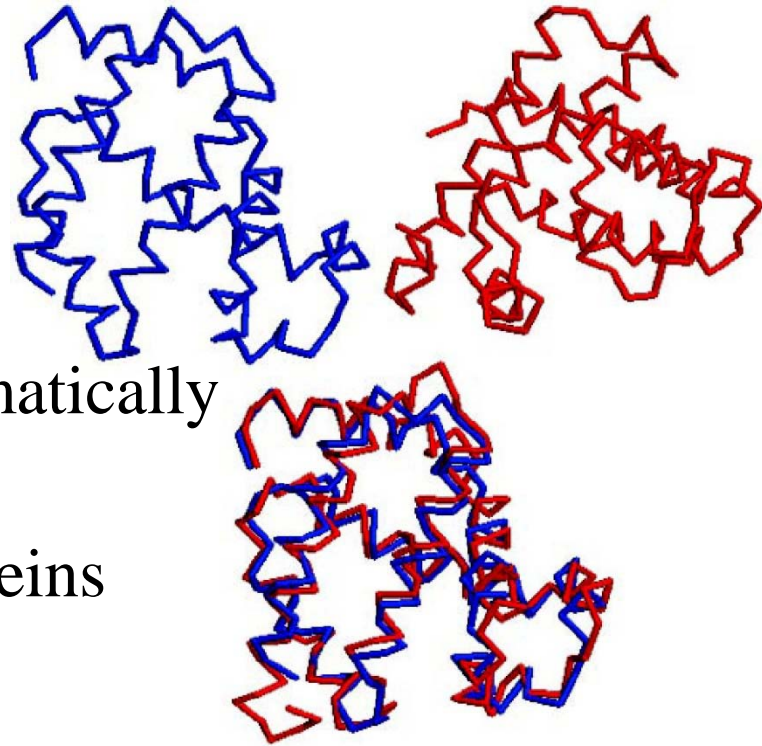
Evolution and Classification

- for very similar proteins, easy
- more remote ?
 - maybe



Forget Evolution

- Is the hierarchy really justified ?
 - at low levels maybe
 - at higher levels ? ($\alpha, \alpha/\beta, \dots$)
- better to discover relationships automatically
- Imagine I can compare arbitrary proteins
- have some measure of similarity
- use this to classify
- Huge problem
 - proteins are different sizes and shapes
 - how to compare ?



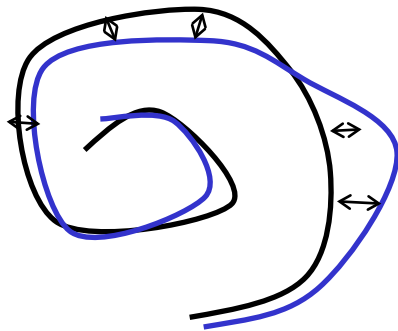
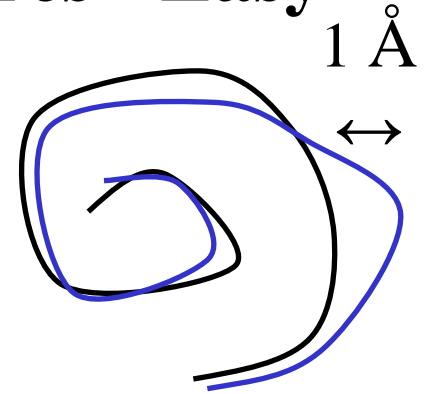
Protein Structure Comparison / Numerical

Most common protein structural question

- how much has my protein moved over a simulation ?
- how similar are these NMR models for a structure ?
- how close is my model to the correct answer ?
- more difficult
 - how similar is rat to human haemoglobin ?
- two cases
 1. same protein, same number of atoms
 2. different proteins
- first
 - measures for easy cases

Numerical Comparison of Structures - Easy

- what units would we like ?
 - scale of similarity (0 to 1.0) ?
 - comparison of angles
 - distance / Å ? most common / easy to interpret
- looks a bit like the average difference between coordinates
- consider analogy with standard deviation / variance



From Standard Deviation to RMSD

Analogy with comparing a set of numbers

- get average (mean) $\bar{x} = N^{-1} \sum_{i=1}^N x_i$

- variance and standard deviation, σ
- apply this to coordinates of r and r'

$$RMSD = \left(N^{-1} \sum_{i=1}^N |\vec{r}_i - \vec{r}'_i|^2 \right)^{1/2}$$

$$\sigma^2 = N^{-1} \sum_{i=1}^N (x_i - \bar{x})^2$$

$$\sigma = \left(N^{-1} \sum_{i=1}^N (x_i - \bar{x})^2 \right)^{1/2}$$

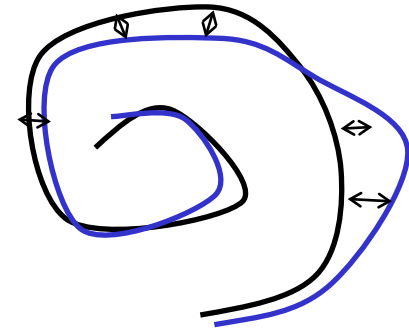
Vital

- formula above, names below
- rms = rmsd = RMSD = root mean square difference

Applying this...

Calculating rmsd

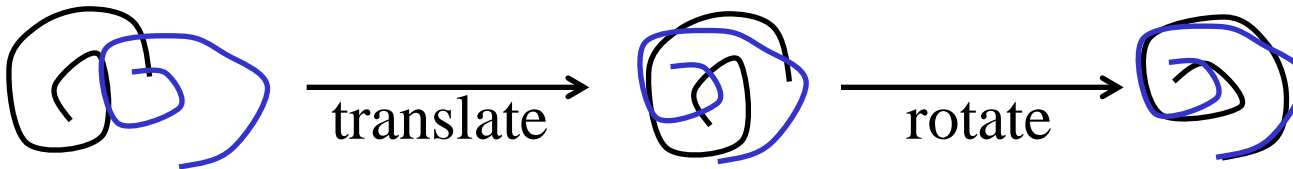
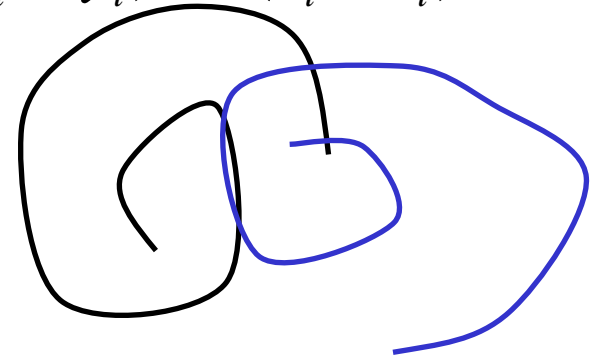
$$RMSD = \left(N^{-1} \sum_{i=1}^N |\vec{r}_i - \vec{r}'_i|^2 \right)^{1/2}$$



- start at one end
- difference between pairs of atoms

$$|\vec{r}_i - \vec{r}'_i|^2 = (x_i - x'_i)^2 + (y_i - y'_i)^2 + (z_i - z'_i)^2$$

- huge problem..
 - coordinates are normally...
- what to do ?



Translation and Rotation

translation

- c.o.m. = centre of mass
- subtract difference vector

$$\vec{r}^{c.o.m.} = \left(\sum_{i=1}^N m_i \right)^{-1} \sum_{i=1}^N \vec{r}_i m_i$$

$$\vec{r}_{diff} = \vec{r}^{c.o.m.} - \vec{r}'^{c.o.m.}$$

- rotation

- messier..

- find rotation matrix to minimise $RMSD = \left(N^{-1} \sum_{i=1}^N |\vec{r}_i - \vec{r}'_i|^2 \right)^{1/2}$

- summary

- translate
 - rotate
 - apply formula
- still not finished

Which Atoms ?

What tells me the shape of a protein ?

- backbone trace

What happens if you include all atoms ?

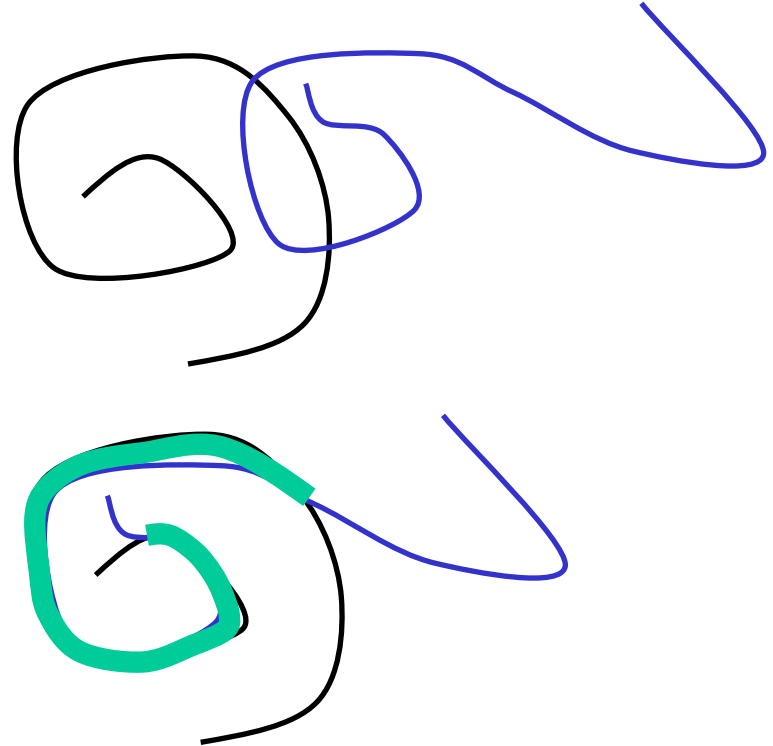
- bigger *rmsd*
- normal choice
 - C^α
- sometimes
 - N, C^α , C
- all atoms ?
 - when a model is very close



Still not finished with simple *rmsd*

Parts Of Proteins

- two models of a molecule
 - mostly very similar
 - is *rmsd* a good measure ?
- identify similar parts



define

```
superimpose ({r},{r'}, {d}) {  
    translate ({r},{r'}, {d})  
    rotate ({r},{r'}, {d})  
}
```

where $\{\mathbf{d}\}$ is some subset of sites

Selection of Interesting Atoms

- define a threshold like `thresh = 2 Å`

```
{d} = { |ri - r'i| } i=1..N  
sort {d}
```

```
diff = rmsd ( {ri}, {r'i} )  
while (diff > thresh) {  
    remove largest d  
    superimpose ( {r}, {r'}, {d} )  
    recalculate distances  
    diff = rmsd ( {r}, {r'}, {d} )  
}  
if (diff < thresh)  
    return {d}, diff  
else  
    return broken
```

- result ? a subset of interesting atoms

Subsets of Atoms

- Originally, quantify structural differences as Å *rmsd*
- Alternative quantity implied
 - number of residues used for *rmsd* below threshold
- implicit rule
 - as number of atoms ↓ calculated *rmsd* ↓

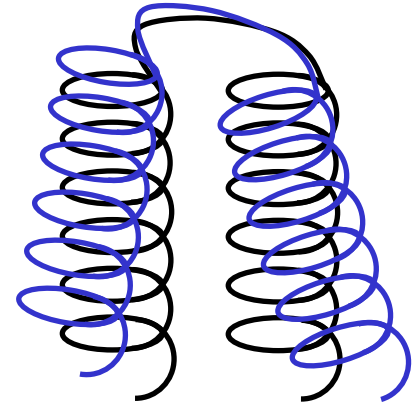
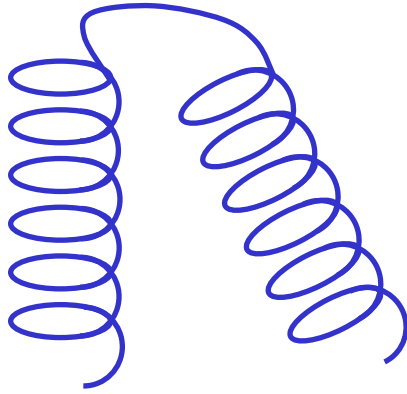
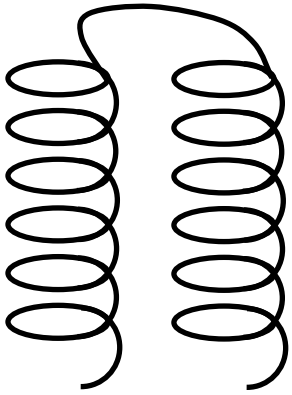
Where we were up to

- Superimposing structures
 - selecting atoms
- Structures of the same size

Where to

- distance matrices for comparison
- finish comparison of structures
 - structures of different sizes
- introduction to modelling
 - interesting bits
- time for some revision next week

Why Not Use *rmsd*



- helices identical, fold identical
rmsd ?
- superposition requires rotation, affects all atoms
- big *rmsd*, but structure has hardly changed
- do not see that helices are identical
- solutions
 - use angles (other problems)
 - distance matrices

Distance Matrices With Numbers

Another characteristic of structures

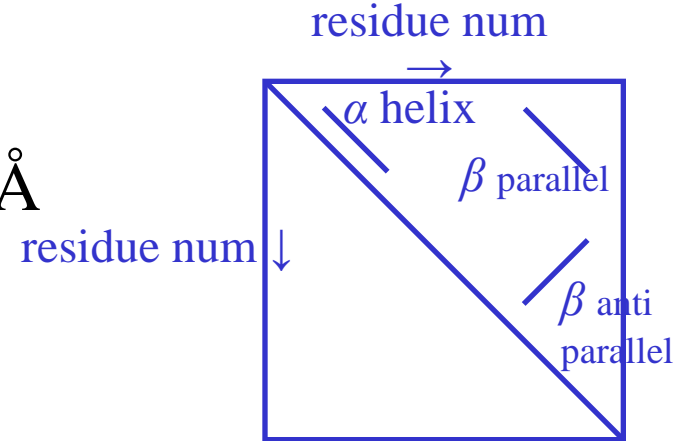
- C^α distance matrices
- simply measure the distance between C^α atoms

	1	2	3	4	5	6	7	...		N
1	0	3.8	6	7	...					
2		0	3.8	5	...					
3			0	3.8	4.5	...				
4				0	3.8					
5					0	3.8				
6						0	3.8			
7							0	3.8		
...								0	3.8	
									0	3.8
N										0

Distance Matrix for Recognising Structure

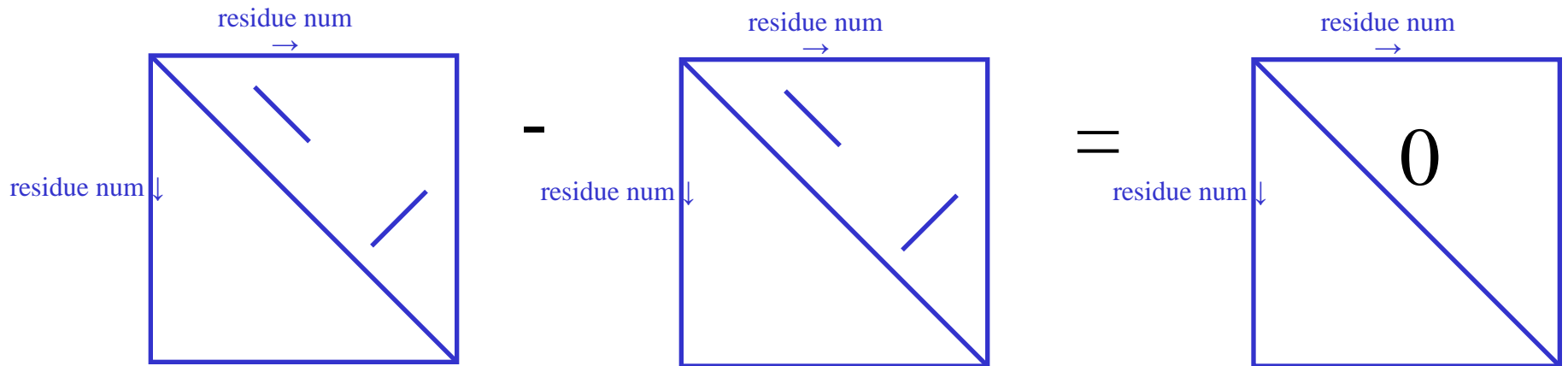
One way to summarise a structure

- plot C^α distance matrix, points below 4 Å
- can make α -helices and β -sheets clear



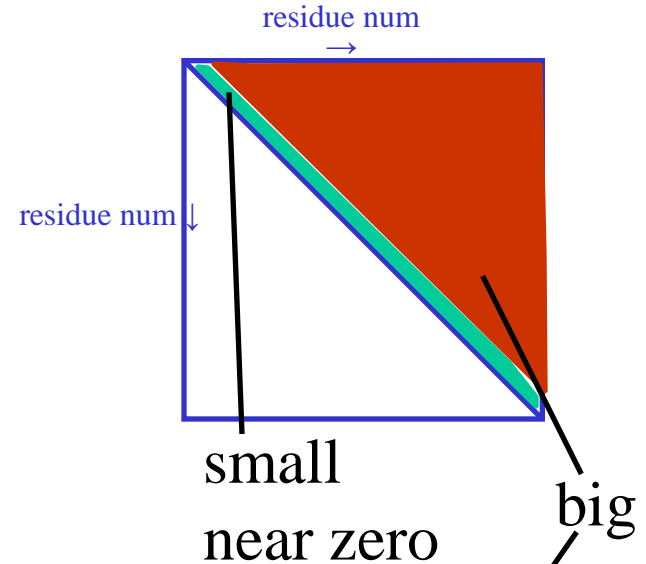
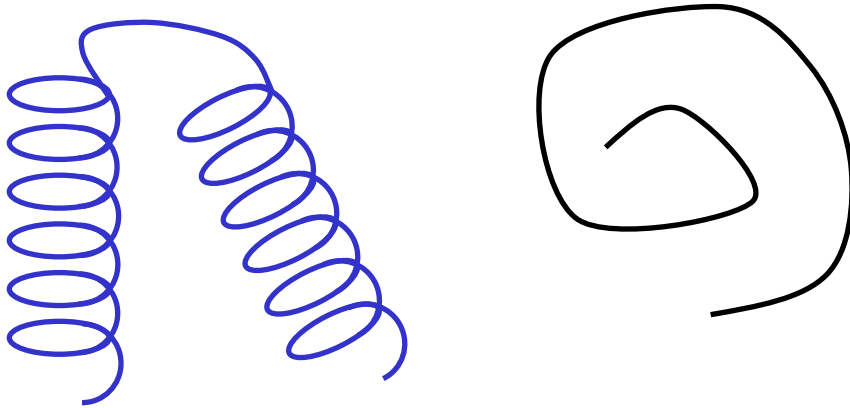
Distance matrix for comparing structures

- take two similar proteins
 - look at the difference of distance matrices

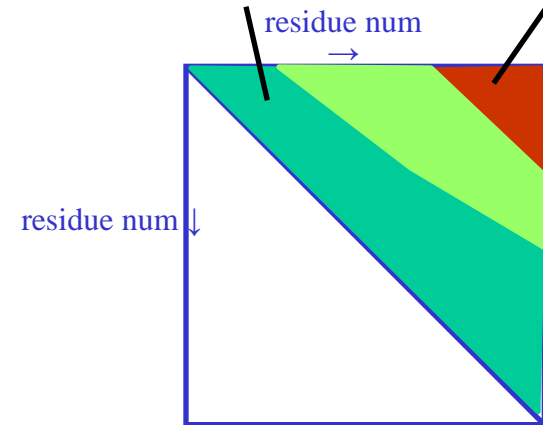
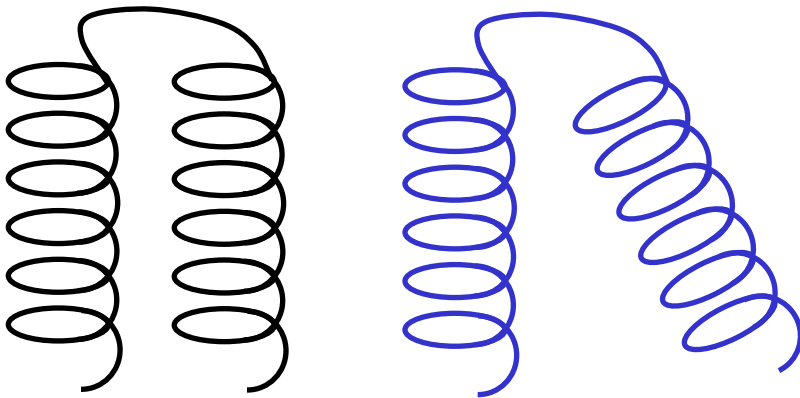


Comparing Distance Matrices

- consider two very different structures



- two related structures



- pictures are better than any single measure, but...

From Distance Matrices to Single Number

For lots of comparisons, single number is more convenient

- root mean square (*rms*) difference of distance matrices
 - define distance between C^α atoms i and j

$$d_{ij} = \left| \vec{r}_i - \vec{r}_j \right|$$

- *rms* of distance matrices measure is

$$rms = \left(\frac{2}{N(N-1)} \sum_{i=1}^N \sum_{j>i}^N (d'_{ij} - d_{ij})^2 \right)^{1/2}$$

- just like all other *rms* quantities
 - normalised over top half of matrix

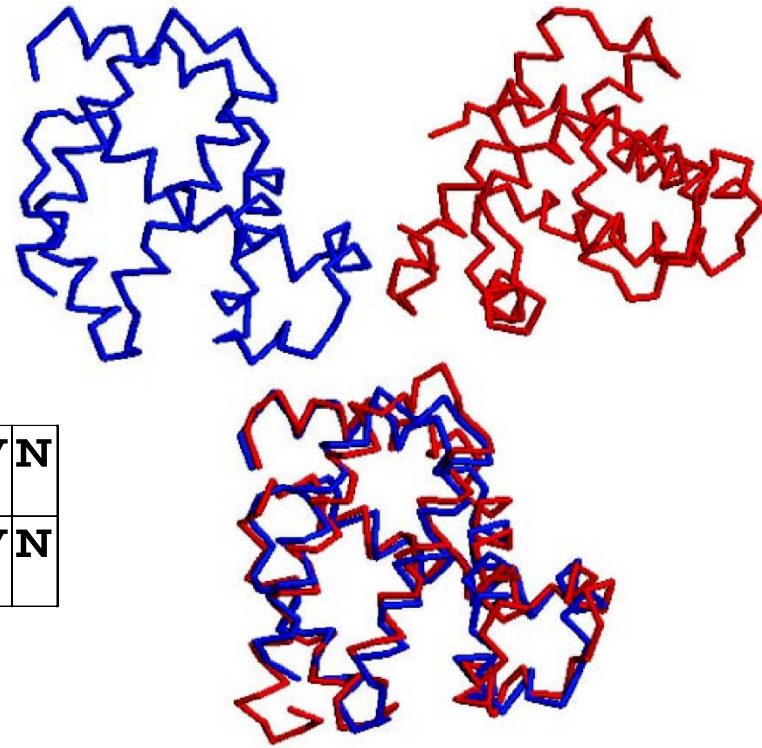
Summary – Comparing Models / Structures

- *rmsd*
 - most popular
 - requires superposition (translate + rotate)
 - can be fooled by "hinge" movements
- to look at the shape of a molecule use C $^{\alpha}$ or backbone atoms
- numbers in Å have a physical meaning
- to look for the common core of a structure, find a subset of backbone
- other measures may be better than *rmsd*
- weakness of all measures
 - a single number can never capture all information

Comparing Different Proteins

- compare red and blue proteins
- if we know which residues match
 - easy (use any *rms* formula)
- which residues match ?
 - sequence alignment ?

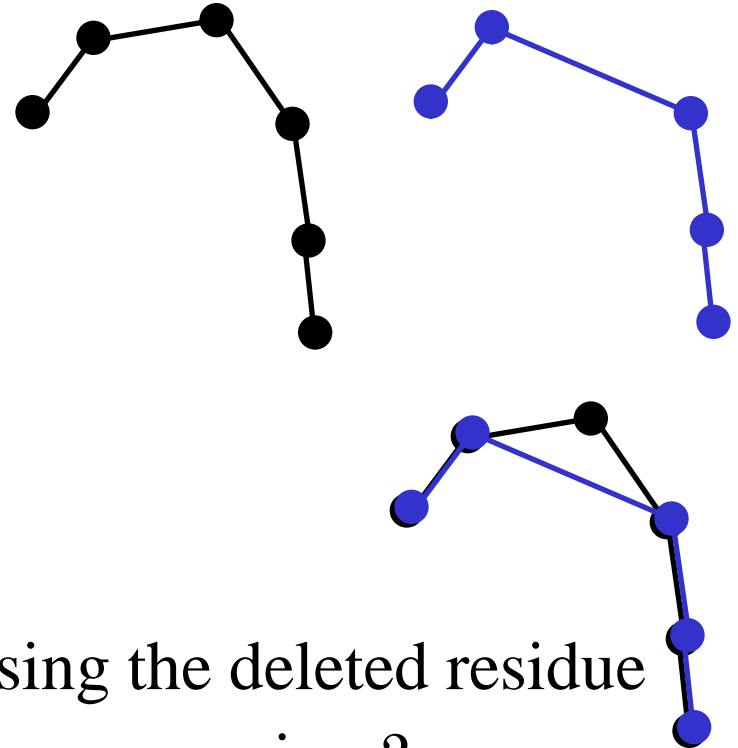
protein 1	A	C	D	W	Y	T	R	P	K	L	H	G	F	D	S	A	C	V	N
protein 2	A	C	D	W	W	T	-	P	K	V	H	G	Y	D	S	A	C	V	N



- green residues - backbone atoms
- pink residues – ignore
- is this useful for similar proteins ? very (rat vs human haemoglobin)
- for very different proteins ? no

Comparing Very Different Proteins

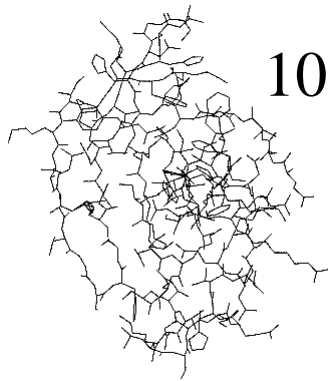
- sequence alignment vs identity
 - as identity ↓, errors ↑
- consequence
 - methods needed
 - operate on C^α
 - do not require sequence
- how difficult ?
 - superposition requires recognising the deleted residue
 - can we use standard dynamic programming ?
 - no
 - gap/insertion at any position, any length
 - combinatorial explosion



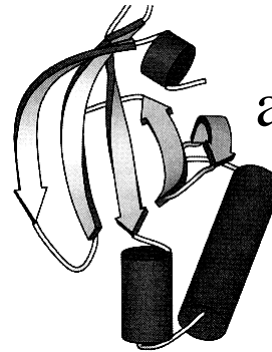
Strategies For Comparing Different Structures

1. use secondary structure

- Combinatorial explosion is the problem
 - reduce size of problem
 - use elements of secondary structure



10^2 - 10^3 atoms

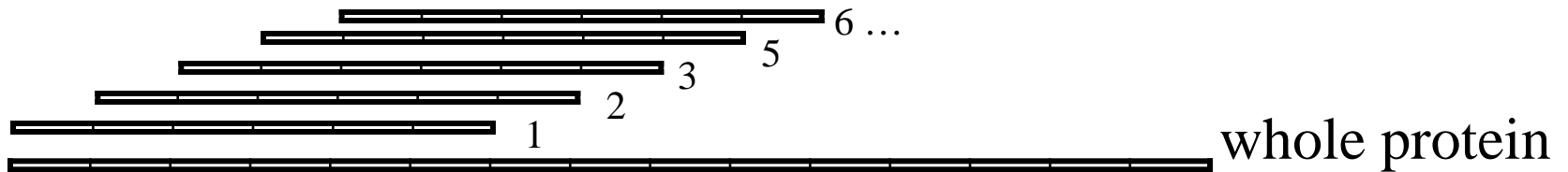


about 8 units

- define secondary structure
- search for superposition
- for each residue
 - find closest C^α in partner structure
 - use the set of matching residues to calculate *rmsd*

2. Peptide fragment strategy

- more general version of idea on previous page
- basis of most popular methods
- Ingredients
 - break protein into overlapping fragments (length 6 or 8)
 - protein is no longer a string of residues nor a whole structure

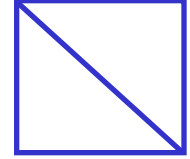
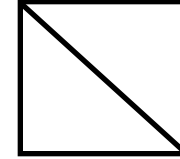


- each fragment is a little distance matrix



Fragment Based Comparison

- any two distance matrices can be compared
- two proteins length N and M can now be compared...



protein 2 fragments ↓

	1	2	3	4	5	...		$N-7$	protein 1 fragments →
1	1.3	1.0	2.0	0.9	...				
2	2.7	2.3	0.5	...					
3	5.5	4.4	...						
4	0.1	0.5	0.3	3.3	4.2	...			
5	1.9	4.4	5.5	0.3	3.3	...			
6	4.4	1.6	1.7	5.0	2.3	...			
...	4.1	3.1	3.3	4.4	0.2	3.3	...		
$M-7$	5.2	1.1	0.1	5.5	4.4	0.1	3.3	0.1	

- imagine *rmsd*
- this is now like a sequence comparison problem

Finding Equivalent Fragments

- find optimal path through matrix
- classic dynamic programming method like sequence comparison

	1	2	3	4	5	...		N-7
1	1.3	1.0	2.0	0.9	...			
2	2.7	2.3	0.5	...				
3	5.5	4.4	...					
4	0.4	0.5	0.3	3.3	4.2	...		
5	1.9	4.4	5.5	0.3	3.3	...		
6	4.4	1.6	1.7	5.0	2.3	...		
...	4.1	3.1	3.3	4.4	0.2	3.3	...	
N-7	5.2	1.1	0.1	5.5	4.4	0.1	3.3	0.1

- like sequence comparison
 - find optimal path through matrix
 - classic dynamic programming method (N & W, S & W)
 - uses gap penalties

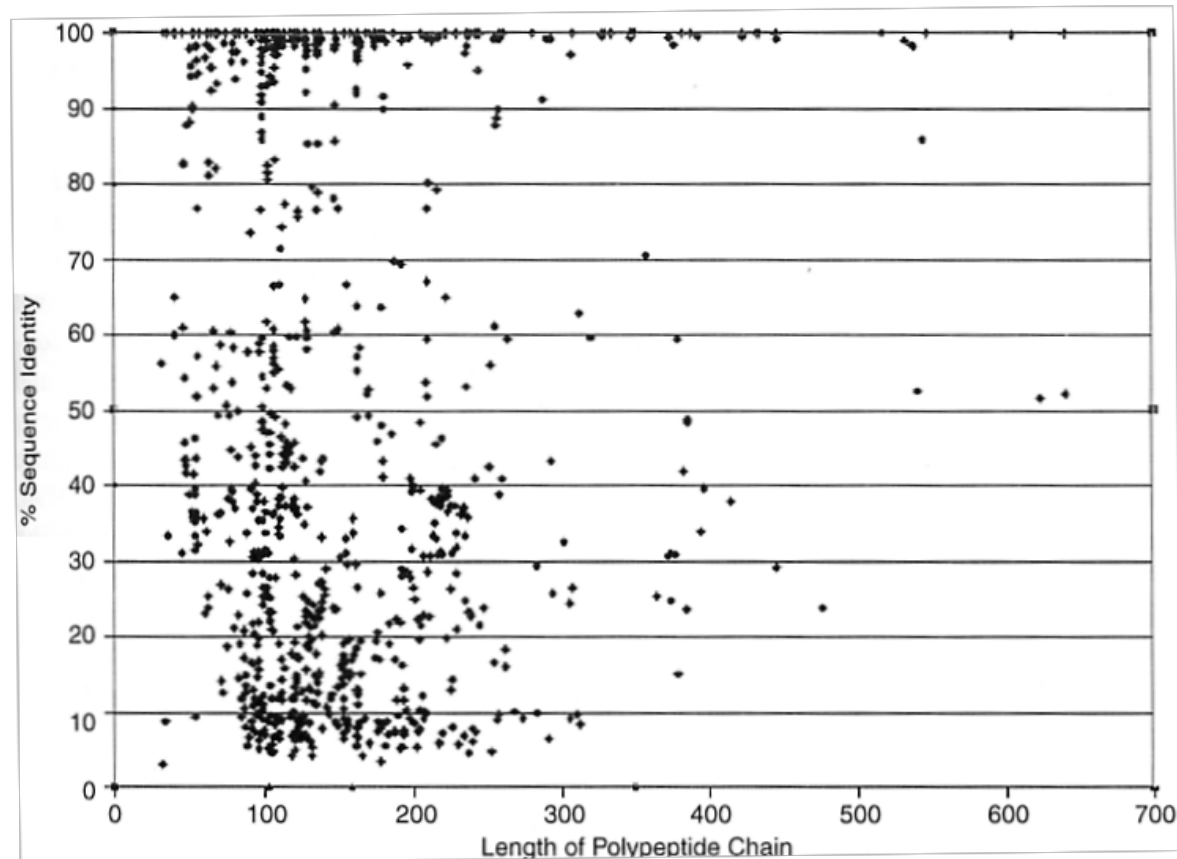
Comparing Different Size Protein Structures

- Break protein into overlapping fragments
- fragments can be compared to each other via distance matrices
- align like sequences
- from aligned fragments, get list of aligned residues
- using aligned residues, calculate *rmsd*, *rms* of overall distance matrices

How Important Are These Similarities ?

- survey 1000 proteins
- find structurally similar pairs
- plot sequence identity

may not be found by
sequence methods



Summary of All Protein Comparisons

Classification of proteins

- could be done by sequence, better by structure

Structure comparison

- for one protein
 - selection of atoms
- for different proteins
 - requires list of matching atoms
- for similar proteins
 - can use pairs from sequence alignment
- for often dissimilar proteins
 - pure structure based method