NMR (Nuclear Magnetic Resonance Spectroscopy)

Literature / background (already in Stine)

- Thomas James chapter http://www.biophysics.org/img/James.T.pdf
- Ferentz, A.E. and Wagner, G., Q. Rev. Biophys, 33, 29-65
 (2000)

current standing

- ≈ 11 % of all current structures solved by NMR
- about 1/4 of smaller structures (<100 residues)

Next 2 Weeks

Background to NMR – chemistry

Calculating structures

- distance geometry
- problems with structures

For chemists: no

- chemical shifts
- 2D and higher
- residual dipole coupling, spin labels
- ...

History

Younger field than X-ray

• 1 ½ Nobel prizes

(Ernst, Wüthrich)

First real protein structure about 1985 or 1986

NMR from our viewpoint

A way to get structures - the focus here

Can provide information on

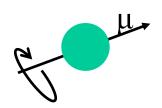
- dynamics, stability
- interactions (other proteins, small molecules)

Overview - how we get coordinates

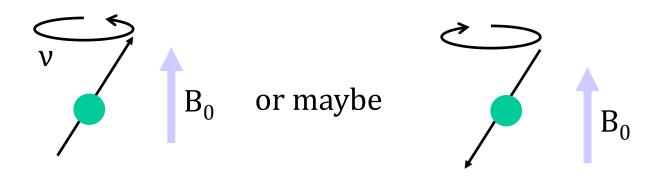
- protein in solution
- record spectra
- assign peaks to ¹H, ¹³C, ¹⁵N nuclei
- record some more spectra
 - distance information (mostly)
 - some internal angles
- reconstruct structure

Nuclei have spin

- have a charge and act like magnets
- put them in a field and they will align with it



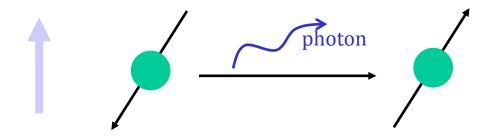
- now apply a magnetic field
 - they "precess" around the field
 - two possible states



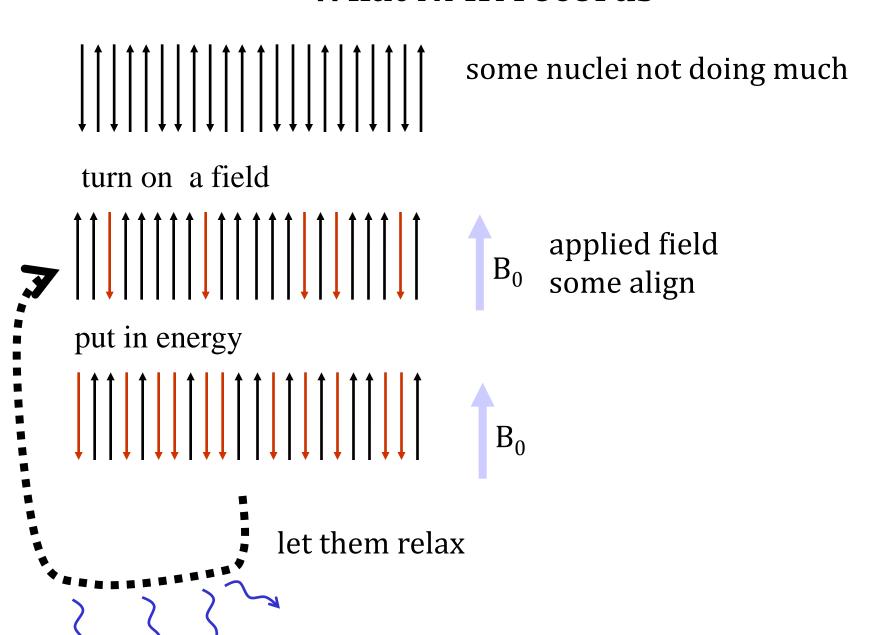
 B_0 is applied field ν speed of rotation (many MHz / 10^6 Hz)

Do nuclei like fighting the field?

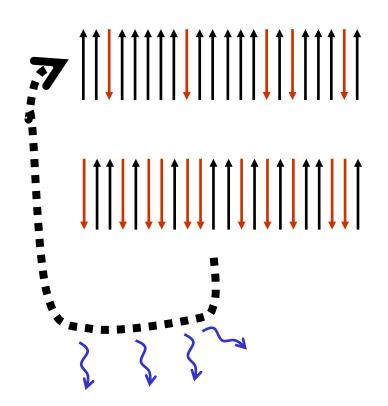
- is a nucleus really happy facing the wrong way?
- what if we push it the wrong way?
 - wants to get to low energy state emits a photon



What NMR records



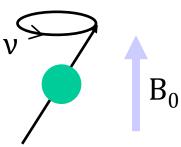
Is this useful?



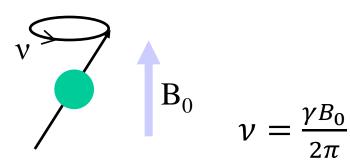
record some photons (radio freq)

• what if the nuclei emit slightly different frequency energy?

- what determines the frequency?
 - energy difference
 - field strength



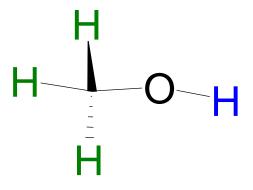
- B₀ applied field
- ν Larmor frequency



 γ magic number for nucleus (gyromagnetic ratio) purely empirical

What is the real field that a nucleus sees?

mixture of outside field and local environment



blue H is different to green H so frequency should change

chemical shift / real spectrum

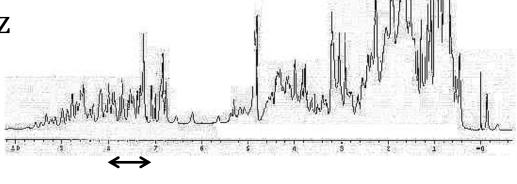
Typical protein

• 100's ¹H

Scales?

• all peaks resonating 100 to 800 MHz (10⁹ Hz)

• whole spectrum 10⁴ Hz



100 Hz

Important nuclei (spin 1/2)

nucleus	sensitivity	notes
¹ H	1	cheap and natural
¹³ C	1.6×10^{-2}	expensive, but only 1% of natural abundance
¹⁵ N	10^{-3}	not cheap, 0.4 % natural abundance
³¹ P	7×10^{-2}	DNA and other PO_4 chemistry, less protein

- but the natural isotopes are ¹²C and ¹⁴N
 - (usually) these isotopes require labelling
- other nuclei? ...

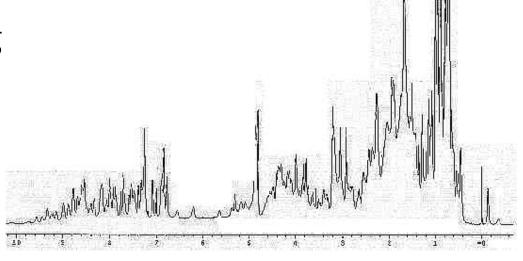
A simple spectrum

Example protein (ubiquitin)

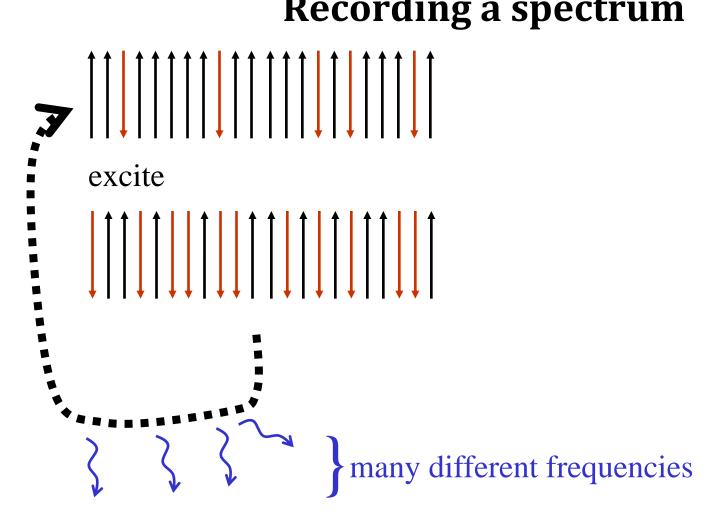
lots of peaks, no structural information yet

Could already

- look at ligand binding
- pk_a of residues



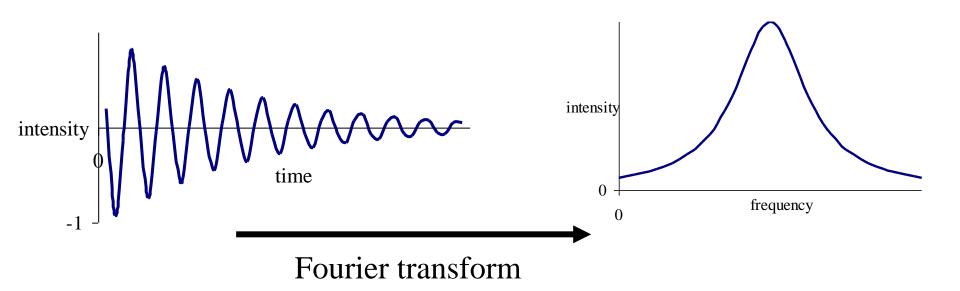
Recording a spectrum



sort out frequencies with Fourier transform

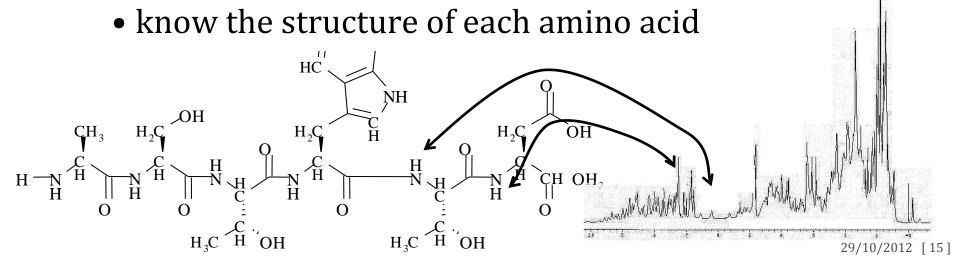
Raw data and Fourier transforms

• raw data will be simple periodic functions + decay



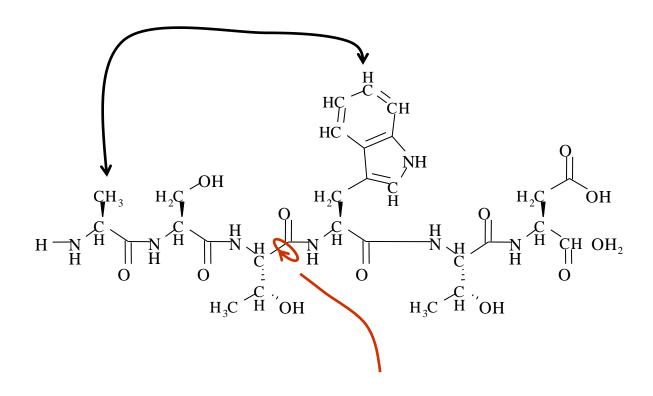
Assignment

- Peaks correspond to ¹H / ¹³C from specific atoms
- structural information given by peaks
 - more later
- Assignment which peaks correspond to which atoms
- Assumption
 - we know exactly which atoms are present
 - sequence of protein known



To calculate structures?

1. distance information



2. dihedral / torsion angle information

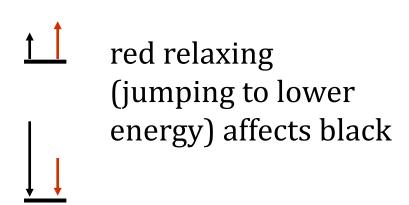
Distance information / the NOE

Most important

- an effect which depends on how close in space nuclei are
- NOE $\propto r^{-6}$
- usually only up to about 5 6 Å

Story

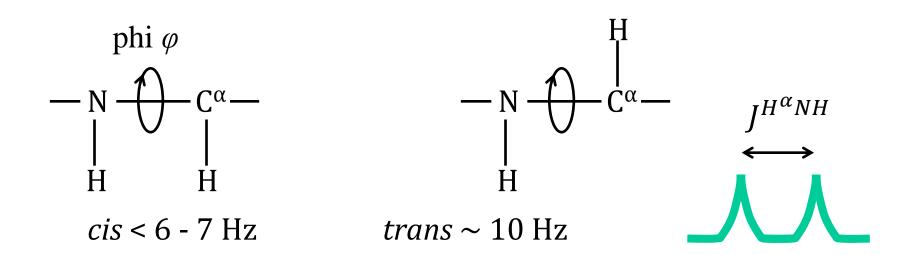
- two spins' dipoles interact
- cross relaxation phenomenon



Other structural information

- NOE information about short (< 5 or 6 Å) distances
- there is more angles
 - mainly *J* coupling

Amide NH to H^{α} coupling

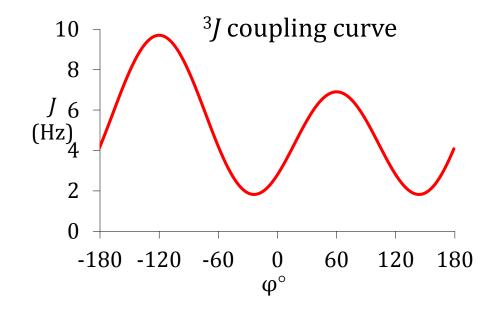


$^{3}J_{HN\alpha}$ coupling

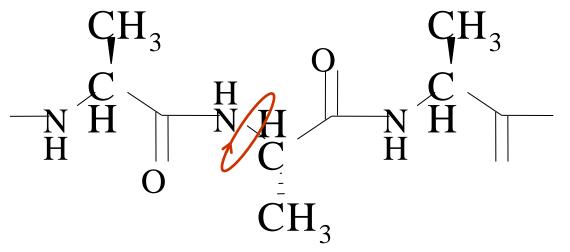
formalised as

$$^{3}J_{HN^{\alpha}} = 6.4\cos^{2}\theta - 1.4\cos\theta + 1.9$$

Problems later



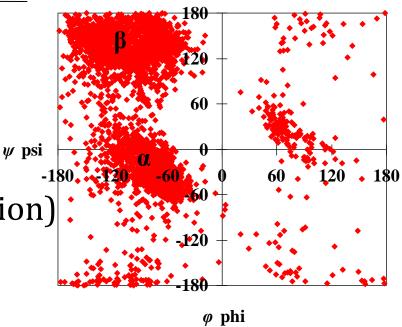
Amide NH to H^α coupling



can help distinguish α from β

• not always seen (exchange / motion)

- NH not always present
- other angles?
 - other vicinal protons
 - C^{α} to C^{β}

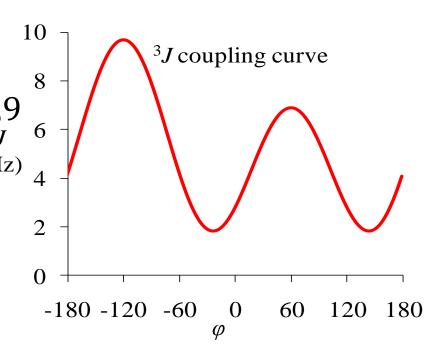


Problems with *J*-coupling

1. we have a formula

$$^{3}J_{HN^{\alpha}} = 6.4\cos^{2}\theta - 1.4\cos\theta + 1.9$$

- most of the time, there is more than one solution
- only use very big J values



2. dynamics & errors in J measurement more serious than they appear! look around -90°

Practical NMR

- We have some basic methods
- Real NMR
 - more techniques
 - 2D and more
 - identifying specific kinds of atom
 - spreading peaks out

Information summary

phenomenon	assignments	structure	
chemical shift	important	not much used	not in Folien
spin-spin (<i>J</i>) coupling	important	torsion angles	
NOE	important	distances	

- more spectroscopy
 - filtering according to chemistry, atom types
 - *n*-dimensional methods
- structural information
 - labels for broadening / shifting peaks
 - orientation of bonds to reference ..

Structures from NMR data

Available information

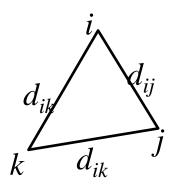
- distances
 - short (5 to 6 Å)
 - incomplete
- some dihedral / torsion angles
- does this define a structure ?
 - strictly no
- Coming
- distances in 2D and 3D
- Distance geometry two versions

Determining distances (ideal)

• 2 points 1 distance

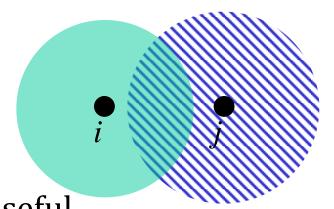
 $\begin{array}{ccc}
\bullet & & \bullet \\
i & d_{ij} & j
\end{array}$

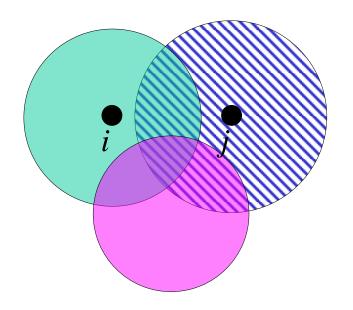
- 3 points 3 distances...
 - think of $3N_{atom}$ distances
 - remember $N_{atom} \approx 10 \text{ or } 20 N_{res}$



Underdetermined distances

- think in terms of triangles ...
 - d_{ik} < 6 Å, d_{jk} < 6 Å
 - where is k?
- a few more distances...
 - more and more distances are useful

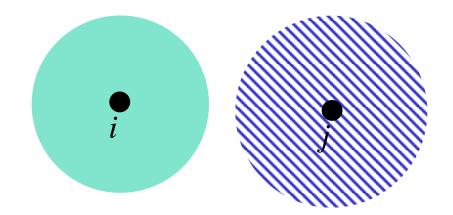




Impossible distances

No overlap?

- experimental error
- nowhere for k to go



Real data

For N residue protein, maybe 5 N_{res} or 10 N_{res}

- want more like $3N_{atom}$ (30 60 N_{res}) distances if perfect
 - needs much more data…
 - lots of chemical data

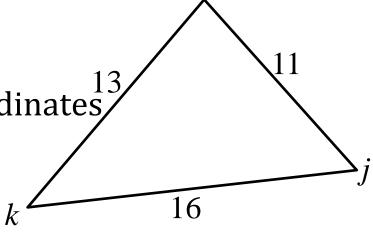
Mission

- gather all experimental data
- mix in chemical data
- make all distance information as tight as possible
- put an upper bound on the distance between every pair of points
- put a lower bound on every distance (less important)
- somehow generate coordinates
- start with toys and triangles

Structures from distance information

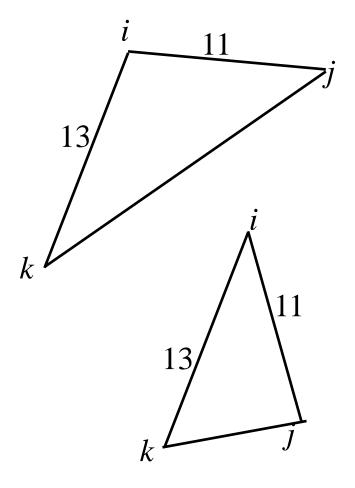
Start in two dimensions...

- ein freundliches Dreieck
 - $d_{ij}=11$ $d_{ik}=13$ $d_{ik}=16$
- fix *i*, put *j* on x-axis and make coordinates
- solve analytically



Underdetermined data

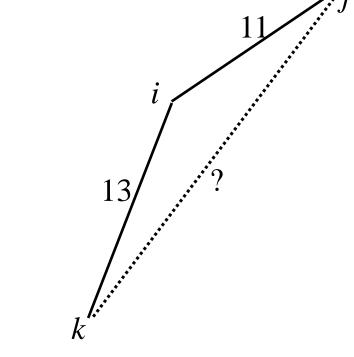
- $d_{ij}=11$ $d_{ik}=13$ $d_{jk}=12-20$
 - more like NMR data
- unique solution?
 - no

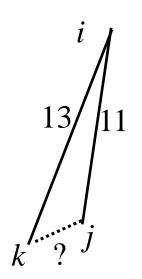


Impossible data

• distance too big $d_{ij}=11$ $d_{ik}=13$ $d_{jk}=25$

- distance too small
 - $d_{ij}=11$ $d_{ik}=13$ $d_{jk}=1$
- no 3D structure





Gathering data

- add in chemistry
- use to get more
 - mix chemistry + measurements
- what comes easily from chemistry?

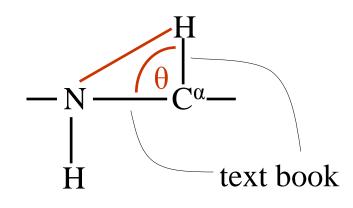
Gather as much data as possible

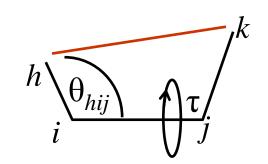
Simple, geometric information

- bonds standard
- angles standard
- simple distances from bond angles
- dihedral / torsion angles

$$d_{hk}^2 = \left(d_{ij} - d_{hi}\cos\theta_{hij} - d_{jk}\cos\theta_{ijk}\right)^2 + \left(d_{hi}\sin\theta_{hij} - d_{jk}\sin\theta_{ijk}\cos\tau_{hijk}\right)^2 + \left(d_{jk}\sin\tau_{hijk}\right)^2$$

- set $\tau = 0$
 - minimum
- \bullet $\tau = \pi$
 - maximum

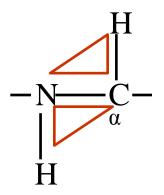




How to get more distance information

- impose some distance limits generally
- intuitively
 - stretch out a protein and there is a limit to length

??



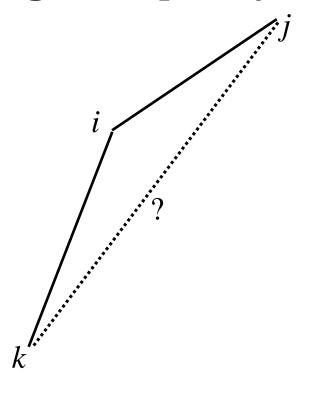
can we formalise this?

More general / triangle inequality

What limits can be worked out?

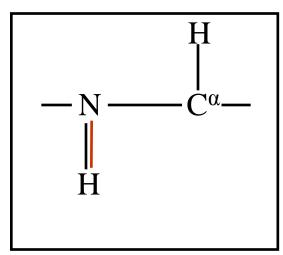
- upper bound $d_{jk} \le d_{ij} + d_{ik}$
- lower bound

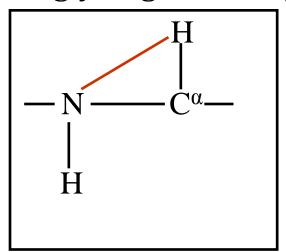
$$d_{jk} \ge \left| d_{ij} - d_{ik} \right|$$

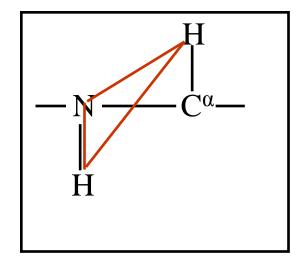


Where to use triangle inequality

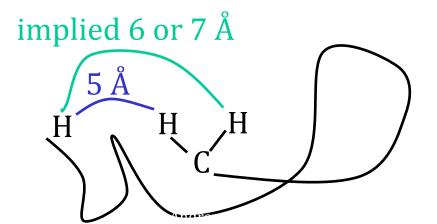
we could avoid some ugly trigonometry





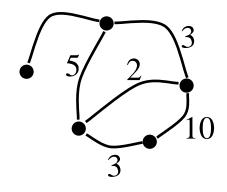


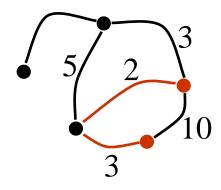
more general



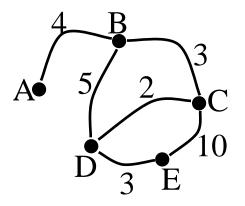
Most general triangle bound inequality

- triangle bound should be satisfied by any three points
- chemists
 - triangle bound smoothing
- informatik
 - all points shortest path problem





All points shortest path (Floyd)

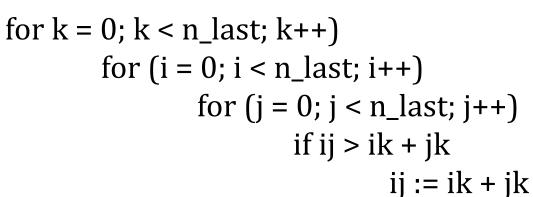


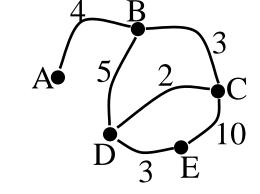
	A	В	C	D	E
A		4			
В			3	5	
C				2	10
D					3
E					

	A	В	C	D	E
A		4	max	max	max
В			3	5	max
C				2	10
D					3
E					

Bound smoothing / Floyd

	A	В	C	D	E
A		4	max	max	max
В			3	5	max
C				2	10
D					3
E					





Running time

• $O(n^3)$

	A	В	C	D	E
A		4	7	9	12
В			3	5	8
C				2	5
D					3
E	And	row Tor			

Distance matrix so far

- we can build a distance matrix of upper limits
 - consistent with all bonds and angles and other information
- can do the same for lower bounds
 - every pair of atoms
 - invent some lower bound (atomic radii)

Does this define a structure?

- almost certainly not
 - still no way to get to a 3D model

From distances to coordinates

How would you build coordinates from distances

- stepwise?
 - error prone, errors add
- history
 - early 80's
 - methods which are tolerant of errors
 - metric matrix method

Metric matrix method

- get best upper bounds
- get best lower bounds
 - guess distances between
 - → trial distance matrix
 - convert to centre of mass matrix (metric matrix)
 - magic conversion to coordinates
 - if metric matrix has three positive eigenvalues
 - error free coordinates
- real coordinates
 - lots of errors
 - initial coordinates not healthy
 - refine

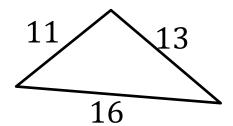
Metric matrix method

- get best lower bounds + upper bounds
 - guess distances between
 - → trial distance matrix
- repeat *n* times
 - get *n* guesses
- some OK, some bad
- repeat until you have 20 or 100 OK structures
- OK = agrees with experimental data + chemically OK

Chirality

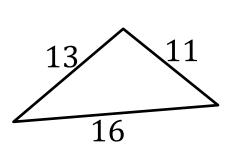
2D version

- can *not* be rotated on to each other
- can not be distinguished by distances



3D

- chirality is random
- problem?no
 - flip all coordinates and check
- local chirality
 - mixture of good and bad
 - difficult to fix



Other distance geometry

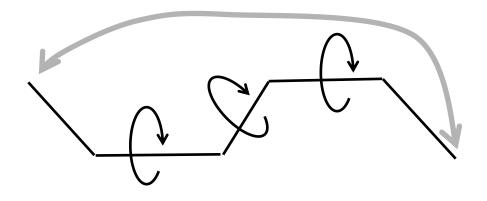
Can we adjust coordinates directly?

Can we work with angles?

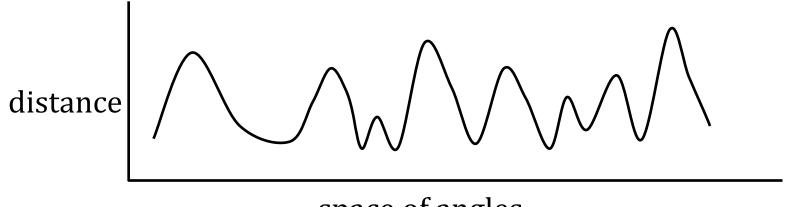
many fewer angles than atoms

Distances and angles

each distance may depend on many included angles

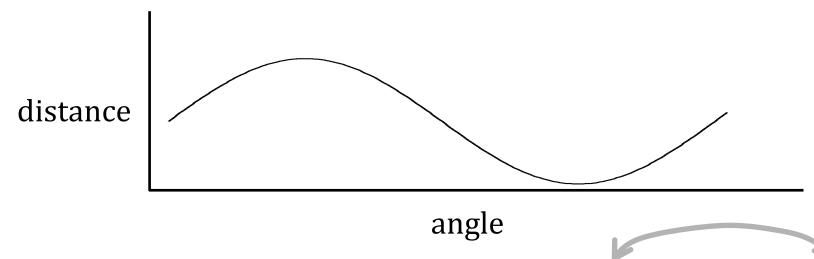


high dimensional space of angles...



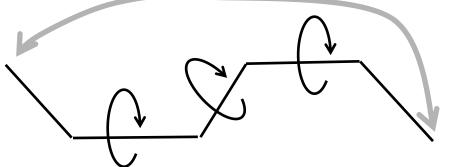
Distances and angles

moving one angle affects one distance simply



- one angle is very important for a
 - short range distance





Optimisation Strategy

Start

- concentrate on distances with few angles in between
- shorter distances become correct

Add in more distances

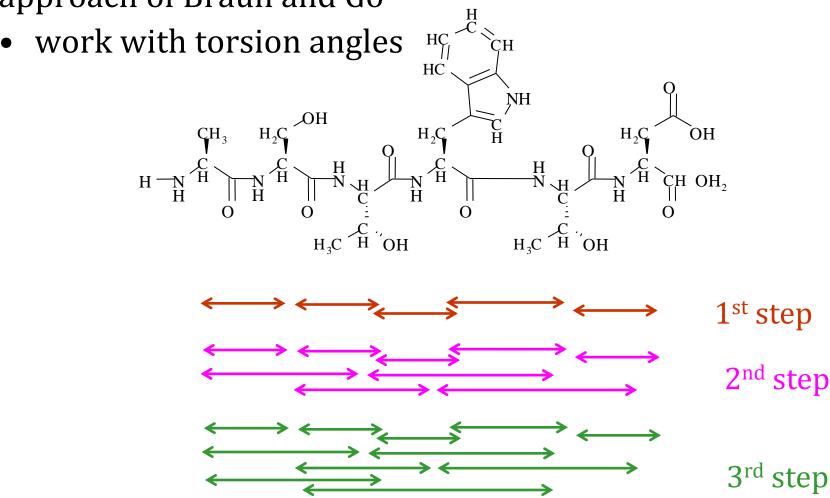
• re-optimise

Add in more distances

• ...

Variable target function

approach of Braun and Gō



Stepwise variable target function method

Collect experimental data

distance	residue	atom	residue	atom	distance
in	1	1	2	2	in space
sequence	;				(Å)
1	5	H^{α}	6	H^N	4.0
0	8	H^{α}	8	H^{γ}	4.4
80	2	H^{α}	82	H^N	4.5
2	3	H^{α}	5	\mathbf{H}^{γ}	5.0
1	7	H^{β}	8	\mathbf{H}^{γ}	3.8
0	3	H^{α}	3	H^N	5.0

• Sort according to distance in sequence

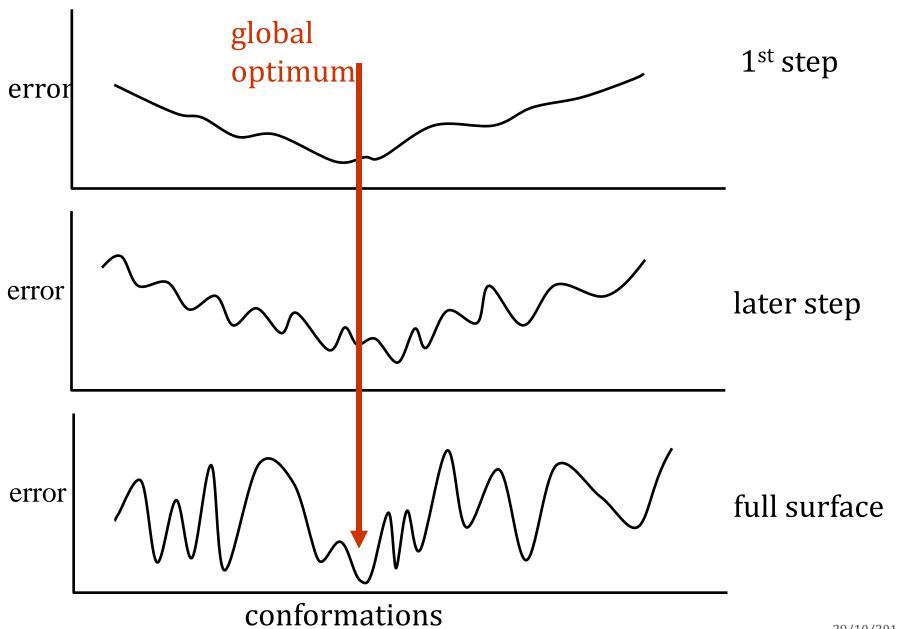
Stepwise variable target function method

distance in sequence	1	atom 1	residue 2	atom 2	distance in space (Å)
0	8	H^{α}	8	\mathbf{H}^{γ}	4.4
0	3	H^{α}	3	H^N	5.0
1	5	H^{α}	6	H^N	4.0
1	7	H^{eta}	8	H^{γ}	3.8
2	3	H^{α}	5	\mathbf{H}^{γ}	5.0
• • •					
80	2	\mathbf{H}^{lpha}	82	H^N	4.5
• • •	•••				

Stepwise variable target function method

distance in sequence	residue 1 e	atom 1	residue 2	atom 2	distance in space (Å)	1 st	2 nd	3 rd .]	later
0	8	H^{lpha}	8	\mathbf{H}^{γ}	4.4	ı	ı	ı		ı
0	3	H^{lpha}	3	H^N	5.0	ļ				
1	5	H^{α}	6	H^N	4.0					
1	7	H^{β}	8	H^{γ}	3.8		ļ			
2	3	H^{α}	5	H^{γ}	5.0			ļ		
•••										
80	2	H^{α}	82	H^N	4.5					1
• • •	• • •									

Hope..



Variable target function vs metric matrix

- metric matrix versus variable target function
 - proponents of both
- variable target function probably more popular
 - no problems with chirality

Real implementations of distance geometry

- not small programs
- Input?
 - list of protein sequence
 - set of distances
- most of code
 - libraries of standard amino acids
 - code to do geometry and work with standard geometries
- other information
 - angle restraints
 - convert to distances for metric matrix
 - natural for variable target function

Output from programs

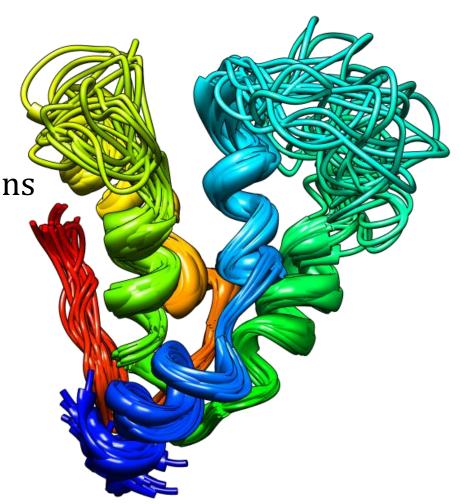
Structure impossible?

- program dies or
- best possible solution

Structure not determined?

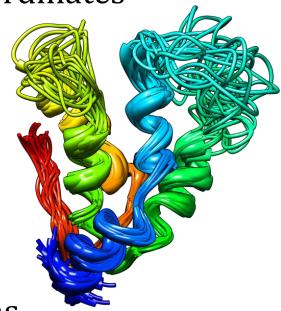
 set of possible conformations (10 to 100)

example 1sm7.pdb



Lots of models in a PDB file

- big difference compared to most x-ray coordinates
- typical
 - ends (C- and N-termini) badly defined
 - loops poorly defined
- spectroscopists say this reflects mobility
- problems with many models
 - difficult to work with
 - arbitrary which to select for calculations
 - averaging usually not a good idea
- Is this the absolute truth? No.
 - number of models arbitrary
 - different methods (programs /details) give different results



Are we finished with making coordinates?

- structures may not be well defined
- can they be improved? probably
 - restrained molecular dynamics (more next semester)
- normal MD $E_{phys}(\vec{r}) = bonds + angles + electrostatics ...$
- restrained MD $E_{total}(\vec{r}) = E_{phys}(\vec{r}) + E_{restr}(\vec{r})$

• and...
$$E_{restr} = \sum_{i} k_i (r_i^{struct} - r_i^{measured})^2$$

- where *i* refers to the distance restraint
- Mission to minimise E_{total}
- result?
- structures
 - agree with restraints + low energy

What else can one do with NMR?

NMR sensitive to dynamics

• is this part of the protein mobile?

Interactions

add small molecule – which parts of spectrum change?

Still more structural information

- residual dipolar coupling
- spin labels

Summary

- What information does one have?
- Is it enough? Is it consistent?
- Two methods to generate structures
- Differences in handling chirality